

Review GLP-1 programs the neurovascular landscape

Bandy Chen,^{1,*} Xiaofei Yu,² Claudia Horvath-Diano,³ María José Ortuño,⁴ Matthias H. Tschöp,^{5,6} Ania M. Jastreboff,³ and Marc Schneeberger^{1,7,*}

¹Laboratory of Neurovascular Control of Homeostasis, Department of Cellular and Molecular Physiology, Yale University School of Medicine, New Haven, CT, USA

²State Key Laboratory of Genetic Engineering, School of Life Sciences, Fudan University, Shanghai, China

³Departments of Medicine (Endocrinology & Metabolism) and Pediatrics (Pediatric Endocrinology), Yale University School of Medicine, New Haven, CT, USA

⁴Department of Genetics, Yale University School of Medicine, New Haven, CT, USA

⁵Helmholtz Zentrum München, Neuherberg, Germany

6Division of Metabolic Diseases, Department of Medicine, Technische Universität München, München, Germany

⁷Wu Tsai Institute for Mind and Brain, Yale University, New Haven, CT, USA

*Correspondence: bac008@health.ucsd.edu (B.C.), marc.schneebergerpane@yale.edu (M.S.)

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SUMMARY

Readily available nutrient-rich foods exploit our inherent drive to overconsume, creating an environment of overnutrition. This transformative setting has led to persistent health issues, such as obesity and metabolic syndrome. The development of glucagon-like peptide-1 receptor (GLP-1R) agonists reveals our ability to pharmacologically manage weight and address metabolic conditions. Obesity is directly linked to chronic low-grade inflammation, connecting our metabolic environment to neurodegenerative diseases. GLP-1R agonism in curbing obesity, achieved by impacting appetite and addressing associated metabolic defects, is revealing additional benefits extending beyond weight loss. Whether GLP-1R agonism directly impacts brain health or does so indirectly through improved metabolic health remains to be elucidated. In exploring the intricate connection between obesity and neurological conditions, recent literature suggests that GLP-1R agonism may have the capacity to shape the neurovascular landscape. Thus, GLP-1R agonism emerges as a promising strategy for addressing the complex interplay between metabolic health and cognitive well-being.

INTRODUCTION

Recent insights into the central regulation of homeostatic feeding have spurred the development of pharmaceutical strategies aimed at effectively managing food intake as an obesity treatment. A pivotal discovery highlights the presence of glucagon-like peptide-1 (GLP-1)-expressing neurons primarily located in the caudal nucleus of the tractus solitarius (NTS) ,¹ crucial for maintaining proper energy balance.² Remarkably, these neurons send numerous ascending projections to hindbrain, midbrain, and forebrain areas, integrating them into both the hedonic and homeostatic control of food intake. $3-6$ Unsurprisingly, GLP-1 receptor (GLP-1R) agonists emerge as the most promising targets for obesity management, prompting the development of numerous receptor agonists such as liraglutide, semaglutide, and tirzepatide. $7-14$ The caveat for exogenous GLP-1 to function as an appetite suppressant is the requirement of a supraphysiological dose with exponentially greater folds in concentration and duration compared to endogenous GLP-1 (Figure 1). Functioning as an incretin hormone, GLP-1 plays a key role in maintaining blood glucose homeostasis by increasing postprandial insulin secretion and reducing glucagon secretion.¹⁵ Endogenous GLP-1 delays gastric emptying, which slows down the rate of glucose absorption and prevents insulin spikes. Beyond this incretin role, GLP-1 signaling significantly contributes to the regulation of diverse behaviors, encompassing meta-

bolic processes and motivated behaviors such as feeding, 3 fluid intake, 16 and drug consumption.¹⁷ Despite the known mediation of these effects by central GLP-1Rs, the precise origins of endogenous GLP-1 responsible for activating these receptors remain a puzzle. This complexity is heightened by GLP-1's production in two distinct locations within the body—peripherally in the gut, 18 released into circulation, and centrally in distinct brain regions, including the NTS and the olfactory bulb.^{19,20} GLP-1 binding onto hypothalamic and hindbrain centers induces satiety, yet the exact mechanisms and pathways through which GLP-1 enacts its effects on the brain remain elusive. Furthermore, the concentration of endogenous GLP-1 secreted and the routes by which it enacts its effects depend on the strength of the stimulus. For example, a normal meal leads to a postprandial level of GLP-1 that triggers the afferent vagus nerve and results in a vagus relay that communicates its overall incretin effect.²¹ This level of plasma GLP-1 does not stimulate receptors on target organs due to its low concentration and short halflife. Conversely, a large meal, or post-bariatric surgery, will result in a higher level of endogenous GLP-1 that acts on both the vagal system and receptors on target organs. Whether postprandial endogenous intestinal GLP-1 can reach the brain to modulate food intake requires further elucidation. It is plausible that a normal meal does not lead to a level of plasma GLP-1 that can reach the brain via systemic circulation, while a large meal leads to a more robust and prolonged release of intestinal GLP-1 that

Figure 1. GLP-1R expression and function on the neuro-glialvascular unit

Endogenous and exogenous GLP-1 (GLP-1R agonist) share overlapping and distinct physiological properties. Both act as incretin hormones to maintain glucose homeostasis and reduce gastric emptying. Exogenous GLP-1 enhances satiety and demonstrates neuroprotection through its anti-inflammatory properties. Whether endogenous GLP-1 can act centrally remains unknown due to its short half-life.

can act on central nodes to regulate satiety. Understanding the interplay between these dual sources (brain and intestine) of endogenous GLP-1 and their respective roles in modulating physiology depending on the stimulus is critical for unraveling the intricate signaling mechanisms associated with GLP-1.

Over the past decades, obesity has surged to pandemic levels in Western societies.²² GLP-1R agonists have gained prominence for delivering a sustained reduction in weight loss (approximately 10%-20%). 23 Individuals with overweight and obesity often exhibit persistent chronic inflammation, both peripherally and centrally. This inflammatory state is intimately linked to a greater risk of developing neurological diseases, connecting obesity-related metabolic syndrome to cognitive decline and neurodegeneration.²⁴ Intriguingly, besides its role in appetite regulation, GLP-1R agonism displays neuroprotective and neurotrophic actions and minimizes neuroinflammation, such as reduction in brain insulin resistance, microglial activation, reactive astrogliosis, and neurodegeneration.^{25,26} This, coupled with GLP-1's ability to act both peripherally and centrally to regulate metabolic health, has prompted investigations into its potential role in addressing extra-metabolic conditions. In addition to the neuroprotective roles, GLP-1R agonism exerts microvascular protection. GLP-1R agonism alleviates retinal vascular leakage and improves brain-retinal-barrier permeability in models of diabetic retinopathy.^{27–29} Given the neurovascular effects on the retina, and the existing parallelism of vascular functions between the retina and the brain, it is logical to extend the focus of GLP-1's roles to extra-metabolic effects on the brain by assessing comprehensively the action of GLP-1 on the neurovascular unit (NVU), where neuronal, vascular, and immune systems actively communicate, coordinated by a diverse group of

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cells mediating the brain-body crosstalk.³⁰ In this review, we discuss the intersection between GLP-1R signaling in metabolic and neurological disorders, exploring its impact on the structure-function relationship of the neuro-glialvascular unit.

GLP-1R SIGNALING ACROSS THE DIFFERENT SYSTEMS OF THE NEURO-GLIAL-VASCULAR UNIT

Our brain constitutes only 2% of total body weight; however, it consumes 20% of the body's energy at rest in the form of glucose and oxygen.³¹ This is achieved via an intricate network of blood vessels that perfuses the brain and ensures a seamless neuronal-vascular crosstalk known as neurovascular coupling (NVC). Such a sophisticated vascular system intricately interacts with its heterogenic pool of cellular components. This dynamic process involves supplying energy substrates while efficiently eliminating metabolic byproducts to uphold the brain's homeostatic equilibrium. The notion of the NVU underscores the physical location where the structural and functional connection between brain cells and the microvasculature occurs to coordinately regulate the brain-body crosstalk. The diverse expression of GLP-1R on the different cell types of the NVU is indicative of its multifaceted roles in overall brain function (Figure 2).

Neurons

Centrally, GLP-1 is primarily produced by neurons in the NTS, with projections to multiple regions, most notably the hypothalamus.³² The role of central GLP-1 in feeding behavior is well established, while its influence on glucose homeostasis requires more elucidation.³³ GLP-1R neurons in the dorsomedial hypothalamus (DMH) lower blood glucose levels by increasing insulin release through an NTS^{GLP-1}-DMH^{GLP-1R}-dorsal motor nucleus of the vagus nerve (DMV)-pancreas pathway.³³ In contrast, GLP-1R neurons in the paraventricular hypothalamus (PVH) suppress food intake via an NTS^{GLP-1}-PVH^{GLP-1R} pathway.³⁴ The divergence in GLP-1R populations in mediating physiological versus pharmacological responses to GLP-1 further highlights the complexity of GLP-1 signaling and introduces the concept of regional and temporal specificity with GLP-1 central nodes responding differently to varying metabolic shifts.³⁵ Beyond appetite regulation and glucose homeostasis, additional neuronal roles for GLP-1R signaling include the regulation of energy expenditure and the ability to modulate the autonomic nervous system, as well as the brain's reward system. In brief, central stimulation of GLP-1R leads to the activation of brown adipose tissue thermogenesis, resulting in weight loss independently of its effects on food intake.³⁶ Additionally, central GLP-1R signaling directly regulates adipocyte lipid metabolism by modulating sympathetic outflow.³⁷ Selective deletion of GLP-1R in the PVH reduces hypothalamic-pituitary-adrenal axis responses to acute and chronic stress.³⁸ GLP-1R acts on the reward circuitry to diminish cocaine-seeking behavior. $39,40$ Such hedonic effects of GLP-1R signaling extend to a less severe form of addiction, such as alcohol use disorder. Treatment

GLP-1R Signaling effects on Neuro-Glial-Vascular Unit

Figure 2. GLP-1R expression and function on the neuro-glial-vascular unit The broad expression of GLP-1R across the different cell types of the NVU works in tandem to enact its multifactorial effect on the brain and body.

with GLP-1R agonists in alcohol-preferring non-human primates reduces voluntary alcohol drinking. 41 In summary, GLP-1R signaling has a broad effect on the brain to regulate wholebody systemic metabolism. Whether these effects are exclusively conveyed by neurons or are partly mediated by other components of the NVU remains to be elucidated.

Glial cells are a heterogenic pool of cells (astrocytes, microglia, and oligodendrocytes) that integrate all aspects of CNS development and formation. During the maturation process of neural circuits, glial cells allow efficient synaptic communication, adapting to changes in plasticity, maintaining internal homeostasis, and regulating the overall network-level activity within the CNS.⁴² Moreover, glial cells are responsible for providing peripheral information into the neuronal network and have a significant impact on whole-body metabolism.⁴³ Though GLP-1R is primarily expressed in neurons, it is also expressed in a variety of glial cells with distinct roles on each cell type.

Astrocytes

Astrocytes are the most abundant glial cells in the brain. Being a diverse group of cells with regional and temporal specificity, the main role of astrocytes is the maintenance of tissue homeostasis at all levels of CNS organization, extending from molecular (regulation of metabolites and neurotransmitters) to organ (maintenance of the blood-brain barrier (BBB) and glymphatic system).⁴⁴ In astrocytes, GLP-1 plays a multifaceted role by inhibiting glucose uptake, promoting fatty acid utilization, and ensuring the maintenance of mitochondrial integrity and function.⁴⁵ The absence of GLP-1R signaling in astrocytes leads to the production of fibroblast growth factor 21, resulting in improved systemic glucose homeostasis and memory formation.⁴⁵ Furthermore, GLP-1 enhances the supportive capacity of astrocytes to neurons by mediating a metabolic shift from oxidative phosphorylation to aerobic glycolysis.⁴⁶ GLP-1induced astrocytic-lactate generation increases neuronal viability as well as dendrite and axon growth. 46 Notably, the activation of GLP-1R in astrocytes within the NTS is implicated in the control of energy balance through the regulation of food intake.⁴⁷ Treatment with GLP-1R agonist liraglutide plays a role in modulating astrocyte polarization by increasing the number of A2 reactive astrocytes, which are crucial for neuronal development, plasticity, and survival. 48 A similar finding is reported in a mouse model of glaucoma, where treatment with GLP-1R agonist NLY01 substantially reduces A1 astrocyte transformation and retinal ganglion cell dealth.⁴⁹ Interestingly, GLP-1 (9-36), a natural cleavage product of GLP-1, binds to insulin-like growth factor 1 receptor and activates the downstream phosphatidylinositol 3-kinase (PI3K)/protein kinase B/AKT pathway in astrocytes during oxygen-glucose deprivation/reoxygenation injury.⁵⁰ In conclusion, GLP-1R signaling in astrocytes regulates both central and peripheral metabolism, extending from energy balance to neuroplasticity.

Microglia

Microglia are the resident immune cells of the CNS, with the most pronounced diversity during CNS development and following disease or injury.⁵¹ Though the primary source of central GLP-1 stems from the NTS, GLP-1-positive cells colocalize with the microglial marker CD11b and are seen in the mouse cortex, indicating a distinct expression of GLP-1 compared to canonical GLP-1-expressing NTS neurons.⁵² In microglia, GLP-1R activation reverses microglial polarization from M1 to M2 subtypes by suppressing AKT and nuclear factor κ B (NF- κ B) phosphorylation, thereby mitigating microgliosis and astrogliosis.^{53,54} This reversal

leads to enhanced neurite complexity and spine morphology in primary cortical neurons.⁵⁴ Additionally, the activation of microglial GLP-1R in the trigeminal nucleus caudalis suppresses the central sensitization of chronic migraine by inhibiting the downstream PI3K/AKT pathway.⁵⁵ This inhibits microglial cell proliferation, morphological changes, and inflammatory cytokine production, further supporting the role of GLP-1 in inducing a quiescence state in microglial cells.⁵⁵ Intriguingly, the neuroprotective effects against microglia-mediated inflammation in neurodegenerative diseases are observed upon the activation of microglial GLP-1R.⁵⁶ In a mouse model of sporadic Parkinson's disease, treatment with NLY01 protects against dopaminergic neuronal loss and motor dysfunction primarily through the inhibition of microglial-mediated conversion of astrocytes to an A1 neurotoxic phenotype.⁵⁷ However, in a 36-week randomized, double-blind, placebo-controlled study, treatment with NLY01 in participants with early untreated Parkinson's disease did not lead to improvements in motor or non-motor features compared with placebo.⁵⁸ It is possible that reduction in microglial activation and astrocytic conversion alone might not alter pathology. Whether modulation of glial activity is more robust in younger participants remains to be elucidated. To conclude, GLP-1R signaling on microglia attenuates neuroinflammation by suppressing the polarization of microglia to a proinflammatory state.

Oligodendrocytes

Myelin is the structure that surrounds individual axons and maintains saltatory impulse propagation.⁵⁹ The insulation provided by myelin not only enhances the speed of electrical conduction but also acts as a protective barrier, shielding axons from potential damage caused by external forces or inflammatory responses. Oligodendrocytes are the CNS glial cells responsible for assembling myelin and providing metabolic support to myelinated axons. Using single-cell RNA sequencing, GLP-1R expression in oligodendrocytes is found in the hypothalamus.⁶⁰ Mature oligodendrocytes (Olig2+PDGFRa) express GLP-1R in the corpus callosum.⁶¹ Whether GLP-1 has a direct effect on oligodendrocyte and myelin homeostasis or an indirect effect by regulating other components of the NVU still requires clarification. Future studies aimed at characterizing GLP-1R expression in oligodendrocytes on a brain-wide scale will allow us to understand the functions of GLP-1R signaling in oligodendrocytes with spatiotemporal specificity and how this signal is integrated with the rest of the NVU.

Endothelial cells

The brain is one of the most highly perfused organs.⁶² Intriguingly, it forms a fundamental structure, the BBB, which selects the size and type of molecules that can access the brain parenchyma. While fundamental for brain function, such a barrier frustrates pharmacological interventions. Importantly, in the context of this review, both GLP-1 and GLP-1R agonists are capable of crossing the BBB.⁶³ Structurally, the brain vascular layer is comprised of endothelial cells, adjacent vascular smooth muscle cells (VSMCs), and pericytes. Central endothelial GLP-1R regulates the uptake of GLP-1 and its analog into the brain parenchyma. 64 GLP-1R agonists prevent tight junction protein degradation, a protective feature for ischemic stroke in middle cerebral artery occlusion (MCAO) and injury models. This process is achieved via binding to endothelial GLP-1R and stabilizing the BBB.⁶⁵ Colocal-

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ization of GLP-1R with endothelial cell marker von Willebrand factor (vWF) is observed on microvessels of the ipsilateral basal cortex after subarachnoid hemorrhage induction.⁶⁶ Moreover, glutamate excitotoxicity plays a vital role in causing neuronal death during ischemic stroke and is implicated in various neurodegenerative disorders. Studies on animal models of stroke show that administering exendin-4 and liraglutide, either before or after cerebral ischemia, can reduce infarct size, alleviate oxidative stress, and enhance endothelial function.⁶⁷ The combination of single-cell RNA-sequencing analysis and immunostaining unveils a high expression of GLP-1R in mouse retinal endothelial cells, which is reduced under diabetic conditions.⁶⁸ Treatment with GLP-1R agonist exendin-4 restores receptor expression and leads to improvements in retinal degeneration, vascular tortuosity, avascular vessels, and vascular integrity. Due to the similarity between the retinal and brain vasculature, this finding begs the question of whether GLP-1R's protective effects on retinal endothelial cells can also be observed in brain endothelial cells in the context of metabolic dysfunction. Whether endothelial cells regulate the uptake of endogenous GLP-1 in the context of a strong stimulus to mediate energy balance remains unknown. Further elucidation of GLP-1R signaling in endothelial cells can assist in the explanation of stimulus-dependent endogenous GLP-1 transport to and signaling in the brain.

Mural cells

Mural cells are the counterparts to endothelial cells, located on the abluminal side of the vasculature.⁶⁹ The advent of novel tools, including high-resolution intravital optical imaging, calcium imaging, and single-cell transcriptome analysis, has allowed the classification of this heterogeneous cell population.⁷⁰ The two broad types of mural cells are VSMCs and pericytes. VSMCs surround 15% of the brain microvessels, primarily arterioles and precapillaries, and express the contractile smooth muscle protein actin. Pericytes, on the other hand, surround 85% of the microvasculature, primarily capillaries, and do not express smooth muscle actin. In the hypothalamus, GLP-1R-expressing mural cells are largely VSMCs, while glucose-dependent insulinotropic polypeptide receptor (GIPR)-expressing mural cells are mostly pericytes.⁶⁰ Additionally, GLP-1Rs are expressed by VSMCs lining cortical arterioles.⁷¹ Importantly, the demonstrated protection accomplished by GLP-1R agonism on ischemic stroke may result from vasodilation, impacting tissue perfusion of the infarcted brain tissue—a mural-mediated effect. Transcriptomic assessment quantifies a significant increase in receptor expression under diabetic conditions.⁷² Activation of GLP-1R enhances pericyte function, restoring vascular integrity and BBB permeability in diabetic conditions. Exendin-4's effects on alleviating diabetes-induced cognitive impairment in rodents can arise from this mechanism. Recent studies outline a protective role of GLP-1 on pericytes in diabetic retinopathy. In diabetic rats, treatment with GLP-1R agonist lixisenatide prevents retinal pericyte loss.⁷³ These discoveries uncover the existence of GLP-1R on retinal pericytes.⁷⁴ Further investigations on GLP-1R signaling in both brain and retinal pericytes will allow the comparison between regional-specific actions of GLP-1R on the BBB and blood-retinal barrier (BRB).

Overall, the role of GLP-1R signaling on central vascular cells remains relatively unknown in comparison to the peripheral vasculature. Similarly, vascular dysfunction in obesity and metabolic

syndrome has been primarily focused on the periphery. In the following sections, we highlight the effects of overnutrition and obesity on the brain microvasculature and propose microvascular dysfunction as a nexus between obesity and neurological diseases.

OBESITY-INDUCED NEUROVASCULAR UNCOUPLING AND MICROVASCULAR DYSFUNCTION

A hallmark of human obesity is chronic exposure to a high-fat diet (HFD), which can trigger neuroinflammation and cognitive decline, posing a risk factor for neurodegeneration. A HFD's impact on the cerebral vasculature includes compromised BBB integrity and neurovascular uncoupling and remodeling. $75-78$ Higher BMI is associated with decreased cerebral perfusion in resting and concentration single-photon emission computed tomography scans.⁷⁹ Young adults with metabolic syndrome exhibit decreased macrovascular and microvascular cerebral blood flow (CBF) due to smaller vessel cross-sectional area and lower mean blood velocity.⁸⁰ The CBF reduction is attributed to a loss of cyclooxygenase (COX) vasodilation, with similarities in cerebrovascular impairments observed in middleaged adults with metabolic syndrome.^{81,82} The reduction in CBF in younger adults with metabolic syndrome mirrors the reduction seen in normal aging for middle-aged adults, indicating that metabolic syndrome accelerates cerebrovascular health deterioration.⁸³ Additionally, the albumin quotient, indicating the ratio of cerebrospinal fluid (CSF) albumin to serum albumin, is higher in patients with type 2 diabetes mellitus (T2DM), positively correlating with CSF biomarkers of angiogenesis and endothelial cell dysfunction such as vascular endothelial growth factor (VEGF).⁸⁴ While it is evident that reduced CBF serves as a marker for vascular dysfunction, it is crucial to emphasize that both an increase and decrease in CBF can indicate a shift in homeostasis and act as markers for a disease state.⁸⁵

The ability of overnutrition to shape the neurovascular landscape raises questions about the reversibility of diet-induced microvascular dysfunction. As for all interventional studies, it is crucial to determine the appropriate age of the subjects and the optimal dose and length of treatment. For example, in juvenile mice, 8 weeks of HFD is insufficient to induce neurovascular impairments; however, alterations are observed after 16 weeks of HFD.⁸⁶ Likewise, 11 weeks of HFD leads to cognitive impairments only in juvenile mice but not in adult mice.⁸⁷ These discrepancies in phenotypes are crucial to avoid any false positives and negatives. With that in consideration, recent studies have demonstrated improvements in diet-induced NVU impairments using pharmacotherapy. Treatment with telmisartan, a common angiotensin II receptor blocker, normalizes diet-induced neurovascular uncoupling and CBF reduction in juvenile mice.⁸⁶ The potential to use GLP-1R agonism as a pharmacological tool to shape the neurovascular landscape is explored in the following sections.

GLP-1R AGONISM REPAIRS METABOLIC-ASSOCIATED NEUROVASCULAR UNCOUPLING AND MICROVASCULAR DYSFUNCTION

The extensive impact of central GLP-1 on feeding behavior, glucose homeostasis, and cognitive function prompts an exploration of the mechanisms through which GLP-1 enacts its extensive neurophysiological effects. This section dives into the impact of GLP-1 signaling on the intricate relationship between the neuro-glial-vascular unit.

Postprandial increases in plasma GLP-1 align with increased regional CBF in the left dorsolateral prefrontal cortex and hypothalmaus.⁸⁸ Under basal and hyperglycemia conditions, GLP-1 (1–37) improves BBB integrity, elevating the expression of the tight junction proteins occludin and claudin-5 via the cyclic AMP/protein kinase A (PKA) pathway in cultured brain microvascular endothelial cells (BMVECs).⁶⁵ In HFD-fed mice, exenatide (a long-acting GLP-1R agonist) mitigates cortical neuroinflammation and behavioral deficits by modulating microglial M2 polarization.⁸⁹ Cultured human astrocytes treated with exenatide exhibit reduced glial fibrillary acidic protein (GFAP) expression in both normo- and hyperglycemic conditions.⁹⁰ Treating diabetic rats with exendin-4 ameliorates functional and structural alterations in the BBB and blood-CSF barrier by increasing protein levels of tight junctions and aquaporins.⁹¹ In patients with T2DM, liraglutide demonstrates cognitive improvement by activating the dorsolateral prefrontal cortex and orbitofrontal cortex brain regions.⁹² Liraglutide treatment also reverses the reduced diameter and functional density of brain capillaries in HFD-fed rats. 93 In streptozotocin (STZ)induced diabetic rats, liraglutide attenuates inflammatory markers in the cerebral microvasculature without impacting blood glucose levels or body weight. 94 In a mixed murine model of Alzheimer's disease (AD) and T2DM, 20 weeks of liraglutide administration reduces vascular damage, brain atrophy, and neuronal loss and alleviates cognitive impairment.⁹⁵ Linagliptin, a dipeptidyl peptidase-4 inhibitor, improves diabetes-induced cerebrovascular dysfunction by reducing endothelin-1 (ET-1) plasma levels and cerebrovascular hyperreactivity.^{96,97} In 12-month HFD-fed mice, linagliptin treatment restores BBB integrity and pericyte coverage and counters the angiogenic effect of T2DM.⁹⁸ Moreover, using diabetic Goto-Kakizaki rats, treatment with linagliptin for 4 weeks restores cerebral perfusion and improves insulin-induced cerebrovascular relaxation and vascular remodeling but does not affect short-term hippocampus-dependent learning.⁹⁹ In this case, the lack of cognitive improvement might be attributed to the duration of intervention. Linagliptin does not cross the BBB and increases GLP-1 levels; therefore, its associated neuroprotection is hypothesized to arise from GLP-1R signaling.¹⁰⁰ Notably, chronic linagliptin treatment demonstrates neuroprotective effects even in mice lacking GLP-1Rs, suggesting a central action for linagliptin beyond its role in incretin regulation.¹⁰¹

In sum, GLP-1R agonism demonstrates promising effects to counteract obesity-induced neurovascular uncoupling and microvascular dysfunction by ameliorating CBF, BBB integrity, and vascular remodeling in humans, rodents, and *in vitro* models (Figure 2). A pilot study assessing the effect of a single dose of exenatide on healthy nondiabetic subjects found no effect on cerebral and peripheral vasculature or on inflammatory biomarkers.¹⁰² This indicates that the effect of GLP-1R agonism on NVC and the cerebral vasculature might require long-term treatment in the context of metabolic shifts to have a clinically relevant effect. Further research is needed to optimize the onset

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and duration of GLP-1R agonism to both prevent and treat an altered microvasculature.

EFFECT OF METABOLIC DYSFUNCTION ON CEREBROVASCULAR RISK AND RECOVERY

Cerebral ischemia-reperfusion injury (CIRI) exacerbates stroke outcomes due to rapid reperfusion, posing a persistent challenge in recovery due to the limited treatment options and a narrow time window for intervention.¹⁰³ The obesity paradox suggests better cardiovascular outcomes for patients with obesity or overweight compared to lean individuals, 104 a notion that is still debated in the cerebrovascular field. This pattern is also seen in AD, where midlife weight gain increases the risk of neurological complications, while late-life weight gain is proposed as a protective factor.¹⁰⁵ Metabolic syndrome prevalence worsens cerebral microvascular rarefaction and endothelial dysfunction induced by CIRI.¹⁰⁶ In rats, early exposure to HFD correlates with impaired NVC and cerebrovascular dysfunction, leading to increased cerebral injury and unfavorable stroke outcomes.¹⁰⁷ Hyperglycemia, an independent risk factor for poor ischemic stroke outcomes, heightens BBB disruption and activates matrix metalloproteinase-9 (MMP-9), a family of extracellular matrix remodeling endopeptidases.^{108,109} Inhibiting MMP-9 activity counteracts HFD-induced cerebrovascular remodeling, reducing hemorrhagic volumes and improving neurological outcomes post-stroke.¹⁰⁹ Hyperglycemia also elevates hypoxiainducible factor 1 alpha (HIF-1a) and VEGF expression, markers of angiogenesis, in brain microvessels after ischemic reperfusion.¹¹⁰ Knocking out endothelial HIF-1 α ameliorates BBB leakage and brain infarction in diabetic mice.¹¹⁰ Hyperbaric oxygen preconditioning attenuates brain infarct and hemorrhagic transformation (HT) by downregulating HIF-1 α and MMP-1 in hyperglycemic MCAO rats.¹¹¹ Weight loss before stroke enhances recovery by normalizing fasting glucose and insulin resistance.¹¹² With T2DM being a major risk factor for stroke development, diabetes treatments addressing cerebrovascular risk and recovery have garnered significant attention. To optimize interventions, understanding spatiotemporal variations in cerebrovascular changes associated with metabolic-induced neurological impairments is essential for targeted interventions.¹¹³

REPURPOSING OF GLP-1 AGONISTS FOR THE TREATMENT OF CEREBROVASCULAR DISEASES

Stroke

Stroke stands as a major contributor to both fatalities and incapacitation, placing a substantial economic burden on Western societies.¹¹⁴ Thrombolysis has been a standard treatment for acute ischemic stroke for a quarter century. However, its efficacy is confined to less than 10% of patients treated within a 4-h window from stroke onset.¹¹⁵ In recent decades, endovascular thrombectomy has emerged as a valuable therapy, showcasing benefits in early recanalization and reperfusion, but its widespread use and enduring effectiveness remain constrained.¹¹⁶ Despite these strides, there persists a need for neuroprotective agents to extend the treatment time window and enhance functional outcomes in ischemic stroke. Diabetes exacerbates the risk of stroke and is implicated in roughly 20% of diabetesrelated deaths, underscoring the interconnected mechanisms of diabetes and stroke. Given the broad usage of GLP-1R agonists for the treatment of obesity and T2DM and its roles in neuroprotection, it offers a promising avenue for future therapeutic breakthroughs in cerebrovascular therapy. In a mouse cerebral ischemia model, there is an increase in GLP-1R expression in the CA1 region after stroke, suggesting a compensatory mechanism for neuronal protection.¹¹⁷ Interestingly, GLP-1R expression exhibits a biphasic response, peaking within 24–48 h after the ischemic insult, followed by a drop, and then a subsequent increase after $1-2$ weeks.¹¹⁷ This biphasic response parallels findings for VEGF-A in response to ischemia.¹¹⁸ However, conflicting reports indicate a decrease in GLP-1R expression at various time points after stroke induction.¹¹⁹ A possible explanation is that GLP-1R expression increases as a compensatory mechanism to the physiological insult and drops back to the homeostatic level after vascular remodeling is established. The parallelism between GLP-1R and VEGF-A expressions further reinforces the link between GLP-1R signaling and cerebral vascular plasticity.

Preclinical studies indicate treatment with GLP-1R agonists as potential complementary interventions to canonical cerebrovascular interventions. Exendin-4 treatment in hyperglycemic mice inhibits MMP-9 activation, reducing infarct growth after cerebral ischemia.¹⁰⁸ In a rat MCAO model, exendin-4 attenuates neurological deficits, brain edema, infarct volume, and BBB permeability, attributed to GLP-1R activation of the Wnt/ β -catenin signaling pathway involved in sprouting and nonsprouting angiogenesis, vasculogenic mimicry, and mosaic vessel formation.^{120,121} This pathway inhibits MMP-9 activation, lowers reactive oxygen species (ROS), and mitigates leukocyte infiltration.¹²⁰ Exendin-4's protective effects extend to cortical arterioles with lasting increases in brain tissue partial pressure of oxygen (pO₂) via modulation of CBF.⁷¹ Warfarin-associated HT after cerebral ischemia is a consequence attributed to increased BBB permeability.¹²² Exendin-4 ameliorates warfarin-associated HT, preserves BBB integrity, and suppresses oxidative DNA damage, lipid peroxidation, microglial activation, and neutrophil infiltration through the inhibition of the PI3K/AKT/ glycogen synthase kinase 3 beta (GSK-3 β) pathway.¹²² Astrocyte-dependent mechanisms mediate exendin-4's ability to preserve BBB integrity, reducing astrocyte-derived VEGF-A and increasing tight junction protein expression.¹²³ Chronic exendin-4 treatment normalizes microvessel density, pericyte coverage, and fibrotic scar formation in MCAO T2DM mice.¹²⁴ In retinal ischemia-reperfusion injury, exendin-4 suppresses BRB breakdown by targeting inflammatory genes (e.g., interleukin-1 beta [IL-1b], tumor necrosis factor alpha [TNF-a], and C-C motif chemokine ligand 2 [CCL2]).¹²⁵ Long-lasting exendin-4-loaded microspheres demonstrate greater improvement in various neurovascular parameters when compared to regular exendin-4, such as cortical CBF, cerebral microcirculation, cognitive deficits, brain edema area, and levels of ROS, aquaporin, and GFAP expression.¹²⁶ In an acute ischemic stroke, liraglutide dose-dependently reduces infarct size.¹²⁷ Proteomics mass spectrometry analysis post-MCAO in mice reveals alterations in oxidative stress, cell growth, apoptosis, and inflammatory response after liraglutide administration.¹²⁸ The neuroprotective effect of liraglutide involves inhibiting

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pyroptosis via the nucleotide-binding domain, leucine-rich-containing family, pyrin domain-containing-3 (NLRP3)/Caspase-1/ IL-1 β pathway.¹²⁹ GLP-1R knockdown abolishes liraglutide's protective effect, implicating nuclear factor erythroid 2-related factor 2 (Nrf2) activation and M2 polarization.¹³⁰ NLRP3 inhibition improves diabetes-mediated cognitive impairment and vascular integrity, preventing the hypoxia-mediated decrease in BDNF (brain-derived neurotrophic factor) secretion.¹³¹ GLP-1 alleviates NLRP3 inflammasome-associated inflammation in perivascular adipose tissue, suggesting a similar interaction between GLP-1 and NLRP3 in the cerebral vasculature.¹³² Of note, BDNF-mediated mitophagy alleviates hyperglycemiainduced BMVEC injury.¹³³ Interestingly, GLP-1R/BDNF/ (tropomyosin receptor kinase B) TrkB signaling modulates hippocampal neuroplasticity in HFD-induced diabetic mice, 134 contributing to GLP-1's neuroprotective effects on cerebrovascular remodeling. $134-136$ In diabetic rats with MCAO, treatment with liraglutide augments Nrf2 and heme oxygenase 1 (HO-1) expression in the cerebral ischemic tissue.¹³⁷ Pretreatment for 2 weeks reduces infarct volume in both diabetic and nondiabetic rats.¹³⁸ while post-treatment enhances VEGF expression without altering cortical CBF.¹³⁹ Delayed administration of liraglutide improves microvessel density and endothelial cell proliferation and upregulates the expression of VEGF.¹⁴⁰ When the administration is extended, liraglutide treatment increases the number of neuronal nuclei, GFAP, vWF, and GLP-1R in the cerebral ischemic area.¹⁴¹ In both delayed scenarios, there is neurovascular remodeling accompanied by improvements in glucose metabolism and neurological function.^{140,141} Both exendin-4 and liraglutide enhance CBF and reduce oxidative stress and cognitive deficits in MCAO diabetic mice.¹⁴² In short, GLP-1's influence on the cerebral vasculature is primarily mediated by its anti-inflammatory properties that prevent detrimental insults from an overactive inflammatory response.

Other GLP-1 analogs also have been demonstrated to reduce ischemic damage after CIRI. Semaglutide, with a longer half-life than liraglutide, demonstrates greater protection against ischemic damage.¹²⁷ Semaglutide treatment reduces inflammatory M1 microglia and A1 astrocytes after ischemic stroke.¹⁴³ In this context, complement (C)3d⁺ A1 astrocytes block BBB permeability in the neuroinflammatory response. The capability of semaglutide to block the astrocyte phenotype conversion suggests that GLP-1R agonists may treat uncontrolled neuroinflammatory-induced neurological disorders¹⁴³ since the extinction of neuroinflammation is complemented by improved growth factor signaling and neurogenesis in hippocampal areas.¹⁴⁴ Immediate and delayed lixisenatide treatment, an analog of exenatide, upregulates VEGF and endothelial nitric oxide synthase (eNOS) expression. This effect is blocked by exendin $(9-39)$.¹⁴⁵ Similarly, lixisenatide administration in diabetic rats reduces cerebral infarct volume, neuronal apoptosis, oxidative stress, and inflammation.^{146,147} Chronic treatment with linagliptin pre- and post-stroke decreases ischemic brain damage in both middleaged diabetic and nondiabetic mice.¹⁴⁸ In the genetically diabetic-obese (*db/db*) mice, chronic post-treatment with linagliptin improves CBF, BBB integrity, and cognitive performance and attenuates cerebral oxidative stress and brain atrophy.¹⁴⁹ The improvement in functional outcome after stroke is attributed to the stromal cell-derived factor 1 alpha (SDF-1a)/C-X-C motif

chemokine receptor 4 (CXCR4) pathway involved in wound healing, angiogenesis, and proliferation.150,151 Using *in vitro* BMVECs, linagliptin ameliorates the lack of proliferative and migratory abilities of BMVECs by enhancing the sirtuin 1 $(SIRT1)/HIF-1\alpha/VEGF$ pathway.¹⁵² Sitagliptin, with a shorter half-life relative to linagliptin, offers protection against CIRI through the GLP-1R-mediated transient recptor potential (TRP)/calcitonin gene-related peptide (CGRP) signaling pathway involved in vasodilation.^{153,154} Of note, overexpression of CGRP protects against hyperglycemia-induced BMVEC damage by suppressing extracellular signal-regulated kinase (ERK)/HIF-1/ VEGF signaling.¹⁵⁵ The pathways mediated by GLP-1R signaling converge into a common theme: anti-inflammatory-mediated cerebral vascular remodeling.

Traumatic brain injury

Compared to CIRI, traumatic brain injury (TBI) due to mechanical impact is caused by different primary insults with similarities in the pathogenesis of these cerebral injuries.¹⁵⁶ TBI induced by controlled cortical impact (CCI) mimics cerebral edema seen in human TBI. Chronic HFD feeding worsens functional outcomes and decreases brain recovery post-TBI by aggravating neuroinflammation and oxidative stress.¹⁵⁷ Higher plasma GLP-1 levels are associated with a greater risk of TBI-induced mortality and may indicate severe central resistance to endogenous GLP-1 in nonsurvivors compared to survivors.¹⁵⁸ Expression of GLP-1R levels decreases significantly after TBI.¹⁵⁹ Exendin-4 restores BBB integrity, reduces neuronal apoptosis, and improves cognitive impairment after mouse TBI induction.¹⁶⁰ A similar effect is seen in rats, with exendin-4 treatment promoting neurological, cognitive, and CBF recovery by attenuating inflammatory responses.¹⁶¹ Impairment of the glymphatic system is a major contributor to the neuropathological changes and cognitive impairment following TBI due to the accumulation of various neurotoxic substances such as amyloid beta and tau protein.¹⁶² Intriguingly, GLP-1R activation in TBI improves glymphatic system dysfunction, alleviating reactive astrogliosis and loss of perivascular aquaporin-4.¹⁶⁰ Post-treatment with liraglutide after CCI improves BBB integrity and sensorimotor function, reduces cerebral edema, and limits cortical tissue loss.^{163,164} Neuroinflammation reduction is evident in lower microglial expression, although astrogliosis remains unaffected, possibly due to the observed time point. 165 Additional research contrasting the effect of GLP-1R agonism on different types of cerebrovascular disease will clarify the overlapping and distinct pathways impacted by GLP-1's anti-inflammatoryinduced vascular plasticity.

Overall, repurposing GLP-1 for cerebrovascular diseases offers an innovative approach to address these challenging health issues (Figure 3). More research is needed to explore the usage of other metabolic drugs, such as sodium/glucose cotransporter 2 (SGLT2) inhibitors and metformin, to lower cerebrovascular risk factors and treat cerebrovascular diseases and their associated comorbidities.

EXTENSION OF NEUROVASCULAR REMODELING TO MYELINATION AND CSF DYNAMICS

The capability of GLP-1 to influence the neurovascular landscape suggests a connection to its role in mediating cognitive

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Figure 3. GLP-1 ameliorates neurovascular uncoupling and promotes cerebrovascular remodeling GLP-1R signaling ameliorates obesity-induced neurovascular uncoupling and cerebrovascular risk through improvements in CBF, BBB integrity, and vascular remodeling. Treatment with GLP-1R agonists in both humans and rodents mitigates glial polarization and neuroinflammation. The induction of an anti-inflammatory state reduces vascular damage and neuronal loss and increases tight junction expression, pericyte coverage, and insulin-induced cerebrovascular relaxation. GLP-1 treatment upregulates the expression of VEGF and induces vascular plasticity in cerebrovascular diseases such as CIRI and TBI.

function, as neurovascular dysfunction is correlated with cognitive impairments in various neurological diseases.¹⁶⁶ Alterations in the brain vasculature correlate with changes in myelin composition, impacting cognitive performance.¹ Chronic overnutrition is associated with lower white matter integrity and cerebral myelin content, ¹⁶⁹ indicating a possible link between obesity and neurological diseases.¹⁷⁰ Consumption of excess HFD leads to oligodendrogliopathy and impedes oligodendrocyte differentiation in the brain and spinal cord.¹⁷¹ HFD-induced demyelination is mediated through astrocyte-linked indirect nicotinamide adenine dinucleotide (NAD⁺)-dependent mechanisms.¹⁷² Inhibition of cluster of differentiation 38 (CD38), an NAD⁺-degrading enzyme, enhances remyelination in regular chow-fed mice and increases astrocytic expression of *Glp1r* and *Igf1*, indicating improved lipid metabolism and insulin signaling.^{45,172} While a minimal effect of CD38 inhibition is seen in HFD-induced demyelination, a combination of CD38 inhibitor and GLP-1R agonist might work in tandem to induce remyelination in obesity. Moreover, caloric restriction promotes remyelination by increasing oligodendrocyte survival and differentiation and decreasing astrogliosis and microgliosis.¹⁷³ Supplementation with nicotinamide, a caloric restriction mimetic, induces myelin production and ameliorates gliosis.¹⁷⁴ The intimate link between the brain vasculature and nutrient consumption on myelin composition indicates that GLP-1 might have an impact on the myelination process.

Myelination

Neurovascular dysfunction with BBB breakdown and reduced CBF is a prominent feature in demyelinating diseases, and therapeutics to modulate the NVU is a potential avenue for preventing demyelination and inducing remyelination.¹⁷⁵ Obesity is a risk factor for multiple sclerosis (MS), and obesity in patients with MS is associated with higher disease severity and a poorer outcome.¹⁷⁶ GLP-1's impact on myelination is evident in preclinical studies where GLP-1R agonists promote remyelination in models of MS. In a cuprizone-induced mouse model of MS, co-treatment with cuprizone and liraglutide for 4 weeks induces remyelination by stimulating oligodendrocyte progenitor cell (OPC) differentiation via anti-inflammatory mechanisms.¹⁷⁷ Cotreatment with cuprizone and NLY01 does not impact demyelination in the corpus callosum.⁶¹ Additionally, post-treatment with NLY01 after cuprizone intoxication does not alter myelin composition or the number of mature oligodendrocytes.⁶¹ Of note, the inflammatory environment and immune trafficking are stable between the vehicle and the treated groups, indicating that posttreatment with NLY01 fails to minimize neuroinflammation and might overshadow its direct impact on oligodendrocytes. In a chronic experimental autoimmune encephalomyelitis (EAE)

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Figure 4. GLP-1 restores CNS remyelination capacity

GLP-1R agonism enhances remyelination by stimulating OPC differentiation. Treatment with GLP-1R agonists inhibits immune cell trafficking into the CNS and attenuates inflammation-induced demyelination.

mouse model of MS, pretreatment with NLY01 delays the onset and attenuates the severity of EAE by inhibiting immune cell trafficking into the CNS. $¹⁷⁸$ The reduction in leukocyte recruitment is</sup> likely attributed to the anti-inflammatory effects of NLY01, leading to improvements in BBB integrity. In symptomatic EAE mice, treatment with exendin-4 leads to remyelination in the lumbar spinal cord.¹⁷⁹ Linagliptin reduces cuprizone-induced demyelination by modulating the adenosine 5' monophosphate-activated protein kinase (AMPK)/SIRT1 and Janus kinase 2 (JAK2)/signal transducer and activator of transcription 3 (STAT3)/NF-kB pathways.¹⁸⁰ The potential of GLP-1R agonism to prevent demyelination or induce remyelination is promising, with further research needed to dissect the role of GLP-1R signaling on oligodendrocytes and OPCs (Figure 4). Whether GLP-1 has a direct effect on myelin composition or indirectly through the alterations of the other components of the NVU remains to be elucidated.

Glymphatic homeostasis

The intricate relationship between GLP-1R signaling and the cerebral vasculature extends to CSF dynamics, crucial for nutrient delivery and waste removal. Neuronal activity-induced functional hyperemia drives CSF dynamics during sleep, and neural activity driven by visual stimulation modulates CSF flow, emphasizing the role of a healthy cerebral vasculature in maintaining cerebral metabolic levels and turnover.^{181,182} Interestingly, dynamic changes in arterial diameter in the absence of neural activation drive perivascular glymphatic CSF inflow and clearance.¹⁸³ Furthermore, alterations in CSF homeostasis and intracranial pressure (ICP) heighten the susceptibility to and are observed in various neurological diseases.¹⁸⁴ In T2DM rats, glymphatic dysfunction contributes to cognitive decline by hindering the clearance of neurotoxic molecules.185,186 Idiopathic intracranial hypertension (IIH), characterized by increased ICP and optic disc swelling, is linked to CSF circulation failure and sinus vein obstruction.^{187,188} Obesity is a major risk factor for IIH development, with greater occurrence in women with obesity.¹⁸⁹ In overweight women with IIH, there is an increase in intracranial CSF volume that accumulates in the extraventricular subarachnoid space with greater venous outflow resistance.¹⁹⁰ In rats, HFD increases CSF secretion without changes in the resistance to CSF drainage compared to the control diet.¹⁹¹ The sexual dimorphic phenotype is observed only in female rats, consistent with the high rates of female patients with IIH and obesity, and demonstrates a 55% increase in ICP.¹⁹² Importantly, weight loss interventions are effective approaches to minimize the risk of IIH and treat patients with IIH.¹⁹³ GLP-1R agonism demonstrates potential in IIH management, with GLP-1R expression in the choroid plexus influencing CSF secretion.¹⁹⁴ In fact, exendin-4 modulates CSF production *in vitro*. ¹⁹⁴ In rodents, this CSF lowering effect translates into exendin-4 reducing ICP in a dose-dependent manner, with effects lasting for 24 h. 194 Intriguingly, clinical studies support the efficacy of GLP-1R agonists in reducing

GLP-1 role on the cerebrospinal fluid

Figure 5. Modulation of glymphatic flux via GLP-1 treatment

Chronic overnutrition increases CSF secretion and ICP, which increases the risk of neurological complications. GLP-1R expression on the choroid plexus functions as a dial that reduces CSF secretion. In human and rodent studies, GLP-1R agonism is capable of lowering CSF secretion and ICP, resulting in improved cognitive function.

ICP, highlighting their potential in treating conditions with elevated ICP. For instance, a case report of IIH caused by Ramadan intermittent fasting led to the hypothesis that a drop in GLP-1 concentration triggers a decrease in GLP-1R activation in the choroid plexus.¹⁹⁵ This results in increased CSF secretion and ICP. Furthermore, in a phase II randomized, double-blind, placebo-controlled trial, exenatide treatment results in clinically meaningful reduction in ICP with improvements in headaches and visual acuity.¹⁹⁶ Unsurprisingly, GLP-1R agonism is now being explored in IIH as a phase III clinical trial to overcome raised ICP (NCT05347147). Lastly, in an open-lab, single-center, casecontrol pilot study, supplementing semaglutide or liraglutide with usual care weight management improves headache frequency compared to usual care weight management alone.¹⁹⁷ In patients with IIH after bariatric surgery, there is an association between a reduction in ICP and an increase in meal-stimulated GLP-1 levels.¹⁹⁸

Together, it is evident that GLP-1R signaling impacts CSF dynamics and ICP (Figure 5). Commonly used off-label ICPlowering drugs, such as acetazolamide, spironolactone, and topiramate, worsen cognitive function. However, treatment with exenatide in a cohort of patients with IIH reduces ICP without affecting cognition.¹⁹⁹ Nevertheless, while these results are encouraging, whether GLP-1R agonism can improve cognitive function through a decrease in ICP for IIH still needs to be validated. Evaluating the mechanistic differences of GLP-1R signaling on various neurological conditions characterized by increased ICP is a critical first step. The capacity of GLP-1 to impact the glymphatic system is currently an early but promising approach to target neurological disorders with glymphatic dysfunction. Being that the glymphatic system is intimately intertwined with the neuro-glial-vascular unit, cerebral vasculature, and myelin composition, the potential of GLP-1 to affect

all these different modalities of the brain due to a positive domino effect highlights GLP-1 as an ideal representative of drug repurposing.

CONCLUSIONS

The intimate link between metabolic and cognitive health sheds light on brain-body communication and redefines certain disorders as neurometabolic. The concept of repurposing antidiabetic drugs for the treatment of neurological diseases is gaining popularity, with metabolic disorders being a major risk factor for neurodegeneration. In recent years, repurposing of GLP-1 mimetics to treat neurological diseases holds promise due to their anti-inflammatory, neuroprotective, and neurotrophic properties. The expression of GLP-1R on diverse cell types and its ability to influence the neurovascular landscape make GLP-1 an ideal candidate to bridge the brain-body and neuro-metabolic crosstalk. Whether GLP-1's effects on these cell types are direct or indirect still requires further clarification, as well as the precise mechanisms by which GLP-1 activates NVC, cerebral vascular remodeling, myelination, and CSF dynamics. A deeper understanding of the pivotal role played by GLP-1R signaling enhances the potential to address both metabolic and neurological disorders, potentially complementing treatments for both types of conditions simultaneously.

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DECLARATION OF INTERESTS

A.M.J. serves on scientific/medical advisory boards for Boehringer Ingelheim, Novo Nordisk, Eli Lilly, Pfizer, Rhythm Pharmaceuticals, WW, and IntelliHealth; serves as a consultant for Scholar Rock; and receives institutional research support from Eli Lilly, Novo Nordisk, and Rhythm Pharmaceuticals. M.H.T. was a Research Cluster Advisory Panel (ReCAP) member of the Novo Nordisk Foundation between 2017 and 2019. He received funding for his research projects by Novo Nordisk (2016–2020) and Sanofi-Aventis (2012–2019). He consulted twice for Boehringer Ingelheim Pharma GmbH & Co. KG (2020 and 2021) and delivered a scientific lecture for Sanofi-Aventis Deutschland GmbH (2020). As CEO and CSO of Helmholtz Munich, he is co-responsible for numerous collaborations of the employees with many companies and institutions worldwide. In this capacity, he discusses potential projects with and has signed/signs contracts for the Helmholtz institute(s) related to research collaborations worldwide, including but not limited to pharmaceutical corporations like Boehringer Ingelheim, Novo Nordisk, Roche Diagnostics, Arbormed, Eli Lilly, SCG Cell Therapy, and others. As the CEO and CSO of Helmholtz Munich, he was/is further overall responsible for commercial technology transfer activities. M.H.T. and A.M.J. confirm that, to the best of their knowledge, none of the above funding sources or collaborations were involved in or influenced the preparation of this manuscript.

REFERENCES

- 1. Larsen, P.J., Tang-Christensen, M., Holst, J.J., and Orskov, C. (1997). Distribution of glucagon-like peptide-1 and other preproglucagon-derived peptides in the rat hypothalamus and brainstem. Neuroscience *77*, 257–270. https://doi.org/10.1016/s0306-4522(96)00434-4.
- 2. Barrera, J.G., Jones, K.R., Herman, J.P., D'Alessio, D.A., Woods, S.C., and Seeley, R.J. (2011). Hyperphagia and increased fat accumulation in two models of chronic CNS glucagon-like peptide-1 loss of function. J. Neurosci. *31*, 3904–3913. https://doi.org/10.1523/JNEUROSCI. 2212-10.2011.
- 3. Turton, M.D., O'Shea, D., Gunn, I., Beak, S.A., Edwards, C.M., Meeran, K., Choi, S.J., Taylor, G.M., Heath, M.M., Lambert, P.D., et al. (1996). A role for glucagon-like peptide-1 in the central regulation of feeding. Nature *379*, 69–72. https://doi.org/10.1038/379069a0.
- 4. Alhadeff, A.L., Rupprecht, L.E., and Hayes, M.R. (2012). GLP-1 neurons in the nucleus of the solitary tract project directly to the ventral tegmental area and nucleus accumbens to control for food intake. Endocrinology *153*, 647–658. https://doi.org/10.1210/en.2011-1443.
- 5. van Bloemendaal, L., IJzerman, R.G., Ten Kulve, J.S., Barkhof, F., Konrad, R.J., Drent, M.L., Veltman, D.J., and Diamant, M. (2014). GLP-1 receptor activation modulates appetite- and reward-related brain areas in humans. Diabetes *63*, 4186–4196. https://doi.org/10.2337/db14-0849.
- 6. López-Ferreras, L., Richard, J.E., Noble, E.E., Eerola, K., Anderberg,
R.H., Olandersson, K., Taing, L., Kanoski, S.E., Hayes, M.R., and Skibicka, K.P. (2018). Lateral hypothalamic GLP-1 receptors are critical for the control of food reinforcement, ingestive behavior and body weight. Mol. Psychiatr. *23*, 1157–1168. https://doi.org/10.1038/mp. 2017.187.
- 7. Jastreboff, A.M., Aronne, L.J., Ahmad, N.N., Wharton, S., Connery, L., Alves, B., Kiyosue, A., Zhang, S., Liu, B., Bunck, M.C., et al. (2022). Tirzepatide Once Weekly for the Treatment of Obesity. N. Engl. J. Med. *387*, 205–216. https://doi.org/10.1056/NEJMoa2206038.
- 8. Garvey, W.T., Frias, J.P., Jastreboff, A.M., le Roux, C.W., Sattar, N., Aizenberg, D., Mao, H., Zhang, S., Ahmad, N.N., Bunck, M.C., et al. (2023). Tirzepatide once weekly for the treatment of obesity in people with type 2 diabetes (SURMOUNT-2): a double-blind, randomised, multicentre, placebo-controlled, phase 3 trial. Lancet *402*, 613–626. https://doi.org/10. 1016/S0140-6736(23)01200-X.
- 9. Pi-Sunyer, X., Astrup, A., Fujioka, K., Greenway, F., Halpern, A., Krempf, M., Lau, D.C.W., le Roux, C.W., Violante Ortiz, R., Jensen, C.B., et al. (2015). A Randomized, Controlled Trial of 3.0 mg of Liraglutide in Weight Management. N. Engl. J. Med. *373*, 11–22. https://doi.org/10.1056/ NEJMoa1411892.
- 10. Rubino, D.M., Greenway, F.L., Khalid, U., O'Neil, P.M., Rosenstock, J., Sørrig, R., Wadden, T.A., Wizert, A., and Garvey, W.T.; STEP 8 Investigators (2022). Effect of Weekly Subcutaneous Semaglutide vs Daily Liraglu-

tide on Body Weight in Adults With Overweight or Obesity Without Diabetes: The STEP 8 Randomized Clinical Trial. JAMA *327*, 138–150. https://doi.org/10.1001/jama.2021.23619.

- 11. Marso, S.P., Daniels, G.H., Brown-Frandsen, K., Kristensen, P., Mann, J.F., Nauck, M.A., Nissen, S.E., Pocock, S., Poulter, N.R., Ravn, L.S., et al. (2016). Liraglutide and Cardiovascular Outcomes in Type 2 Diabetes. N. Engl. J. Med. *375*, 311–322. https://doi.org/10.1056/ NEJMoa1603827.
- 12. Kelly, A.S., Auerbach, P., Barrientos-Perez, M., Gies, I., Hale, P.M., Marcus, C., Mastrandrea, L.D., Prabhu, N., and Arslanian, S.; NN8022-4180 Trial Investigators (2020). A Randomized, Controlled Trial of Liraglutide for Adolescents with Obesity. N. Engl. J. Med. *382*, 2117–2128. https:// doi.org/10.1056/NEJMoa1916038.
- 13. Wilding, J.P.H., Batterham, R.L., Calanna, S., Davies, M., Van Gaal, L.F., Lingvay, I., McGowan, B.M., Rosenstock, J., Tran, M.T.D., Wadden, T.A., et al. (2021). Once-Weekly Semaglutide in Adults with Overweight or Obesity. N. Engl. J. Med. *384*, 989–1002. https://doi.org/10.1056/ NEJMoa2032183.
- 14. Lincoff, A.M., Brown-Frandsen, K., Colhoun, H.M., Deanfield, J., Emerson, S.S., Esbjerg, S., Hardt-Lindberg, S., Hovingh, G.K., Kahn, S.E., Kushner, R.F., et al. (2023). Semaglutide and Cardiovascular Outcomes in Obesity without Diabetes. N. Engl. J. Med. *389*, 2221–2232. https:// doi.org/10.1056/NEJMoa2307563.
- 15. Andersen, A., Lund, A., Knop, F.K., and Vilsbøll, T. (2018). Glucagon-like peptide 1 in health and disease. Nat. Rev. Endocrinol. *14*, 390–403. https://doi.org/10.1038/s41574-018-0016-2.
- 16. Brakey, D.J., Schatz, K.C., Paul, M.J., and Daniels, D. (2023). The role of glucagon-like peptide-1 (GLP-1) in fluid and food intakes in vasopressindeficient Brattleboro rats. Physiol. Behav. *262*, 114093. https://doi.org/ 10.1016/j.physbeh.2023.114093.
- 17. Aranäs, C., Blid Sköldheden, S., and Jerlhag, E. (2023). Antismoking agents do not contribute synergistically to semaglutide's ability to reduce alcohol intake in rats. Front. Pharmacol. *14*, 1180512. https://doi.org/10. 3389/fphar.2023.1180512.
- 18. Duca, F.A., Waise, T.M.Z., Peppler, W.T., and Lam, T.K.T. (2021). The metabolic impact of small intestinal nutrient sensing. Nat. Commun. *12*, 903. https://doi.org/10.1038/s41467-021-21235-y.
- 19. Brierley, D.I., Holt, M.K., Singh, A., de Araujo, A., McDougle, M., Vergara, M., Afaghani, M.H., Lee, S.J., Scott, K., Maske, C., et al. (2021). Central and peripheral GLP-1 systems independently suppress eating. Nat. Metab. *3*, 258–273. https://doi.org/10.1038/s42255-021-00344-4.
- 20. Montaner, M., Denom, J., Jiang, W., Magnan, C., Trapp, S., and Gurden, H. (2023). The local GLP-1 system in the olfactory bulb is required for odor-evoked cephalic phase of insulin release in mice. Mol. Metabol. *73*, 101738. https://doi.org/10.1016/j.molmet.2023.101738.
- 21. Smits, M.M., and Holst, J.J. (2023). Endogenous glucagon-like peptide (GLP)-1 as alternative for GLP-1 receptor agonists: Could this work and how? Diabetes. Metab. Res. Rev. *39*, e3699. https://doi.org/10. 1002/dmrr.3699.
- 22. Kopp, W. (2019). How Western Diet And Lifestyle Drive The Pandemic Of Obesity And Civilization Diseases. Diabetes Metab. Syndr. Obes. *12*, 2221–2236. https://doi.org/10.2147/DMSO.S216791.
- 23. Jastreboff, A.M., and Kushner, R.F. (2023). New Frontiers in Obesity Treatment: GLP-1 and Nascent Nutrient-Stimulated Hormone-Based Therapeutics. Annu. Rev. Med. *74*, 125–139. https://doi.org/10.1146/annurev-med-043021-014919.
- 24. Morys, F., Dadar, M., and Dagher, A. (2021). Association Between Midlife Obesity and Its Metabolic Consequences, Cerebrovascular Disease, and Cognitive Decline. J. Clin. Endocrinol. Metab. *106*, e4260–e4274. https:// doi.org/10.1210/clinem/dgab135.
- 25. Wong, C.K., McLean, B.A., Baggio, L.L., Koehler, J.A., Hammoud, R., Rittig, N., Yabut, J.M., Seeley, R.J., Brown, T.J., and Drucker, D.J. (2024). Central glucagon-like peptide 1 receptor activation inhibits Tolllike receptor agonist-induced inflammation. Cell Metabol. *36*, 130– 143.e5. https://doi.org/10.1016/j.cmet.2023.11.009.

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- 26. Kopp, K.O., Glotfelty, E.J., Li, Y., and Greig, N.H. (2022). Glucagon-like peptide-1 (GLP-1) receptor agonists and neuroinflammation: Implications for neurodegenerative disease treatment. Pharmacol. Res. *186*, 106550. https://doi.org/10.1016/j.phrs.2022.106550.
- 27. Wei, L., Mo, W., Lan, S., Yang, H., Huang, Z., Liang, X., Li, L., Xian, J., Xie, X., Qin, Y., et al. (2022). GLP-1 RA Improves Diabetic Retinopathy by Protecting the Blood-Retinal Barrier through GLP-1R-ROCK-p-MLC Signaling Pathway. J. Diabetes Res. *2022*, 1861940. https://doi.org/10. 1155/2022/1861940.
- 28. Fan, Y., Liu, K., Wang, Q., Ruan, Y., Ye, W., and Zhang, Y. (2014). Exendin-4 alleviates retinal vascular leakage by protecting the blood-retinal barrier and reducing retinal vascular permeability in diabetic Goto-Kakizaki rats. Exp. Eye Res. *127*, 104–116. https://doi.org/10.1016/j. exer.2014.05.004.
- 29. Liu, J., Wang, H., and Huang, C. (2023). Exendin-4, a GLP-1 receptor agonist, suppresses diabetic retinopathy *in vivo* and *in vitro*. Arch. Physiol. Biochem. *x*, 1–10. https://doi.org/10.1080/13813455.2023.2274279.
- 30. Chen, B., Meseguer, D., Renier, N., and Schneeberger, M. (2023). Dynamic rewiring of neurovasculature in health and disease. Trends Mol. Med. *29*, 786–788. https://doi.org/10.1016/j.molmed.2023.06.011.
- 31. Mergenthaler, P., Lindauer, U., Dienel, G.A., and Meisel, A. (2013). Sugar for the brain: the role of glucose in physiological and pathological brain function. Trends Neurosci. *36*, 587–597. https://doi.org/10.1016/j.tins. 2013.07.001.
- 32. Baggio, L.L., and Drucker, D.J. (2014). Glucagon-like peptide-1 receptors in the brain: controlling food intake and body weight. J. Clin. Invest. *124*, 4223–4226. https://doi.org/10.1172/JCI78371.
- 33. Huang, Z., Liu, L., Zhang, J., Conde, K., Phansalkar, J., Li, Z., Yao, L., Xu, Z., Wang, W., Zhou, J., et al. (2022). Glucose-sensing glucagon-like peptide-1 receptor neurons in the dorsomedial hypothalamus regulate glucose metabolism. Sci. Adv. *8*, eabn5345. https://doi.org/10.1126/ sciadv.abn5345.
- 34. Liu, J., Conde, K., Zhang, P., Lilascharoen, V., Xu, Z., Lim, B.K., Seeley, R.J., Zhu, J.J., Scott, M.M., and Pang, Z.P. (2017). Enhanced AMPA Receptor Trafficking Mediates the Anorexigenic Effect of Endogenous Glucagon-like Peptide-1 in the Paraventricular Hypothalamus. Neuron *96*, 897–909.e5. https://doi.org/10.1016/j.neuron.2017.09.042.
- 35. Varin, E.M., Mulvihill, E.E., Baggio, L.L., Koehler, J.A., Cao, X., Seeley, R.J., and Drucker, D.J. (2019). Distinct Neural Sites of GLP-1R Expression Mediate Physiological versus Pharmacological Control of Incretin Action. Cell Rep. 27, 3371-3384.e3. https://doi.org/10.1016/j.celr 2019.05.055.
- 36. Beiroa, D., Imbernon, M., Gallego, R., Senra, A., Herranz, D., Villarroya, F., Serrano, M., Fernø, J., Salvador, J., Escalada, J., et al. (2014). GLP-1 agonism stimulates brown adipose tissue thermogenesis and browning through hypothalamic AMPK. Diabetes *63*, 3346–3358. https://doi.org/ 10.2337/db14-0302.
- 37. Nogueiras, R., Pérez-Tilve, D., Veyrat-Durebex, C., Morgan, D.A., Varela,
L., Haynes, W.G., Patterson, J.T., Disse, E., Pfluger, P.T., López, M., et al. (2009). Direct control of peripheral lipid deposition by CNS GLP-1 receptor signaling is mediated by the sympathetic nervous system and blunted in diet-induced obesity. J. Neurosci. *29*, 5916–5925. https://doi.org/10. 1523/JNEUROSCI.5977-08.2009.
- 38. Ghosal, S., Packard, A.E.B., Mahbod, P., McKlveen, J.M., Seeley, R.J., Myers, B., Ulrich-Lai, Y., Smith, E.P., D'Alessio, D.A., and Herman, J.P. (2017). Disruption of Glucagon-Like Peptide 1 Signaling in Sim1 Neurons Reduces Physiological and Behavioral Reactivity to Acute and Chronic Stress. J. Neurosci. *37*, 184–193. https://doi.org/10.1523/JNEUROSCI. 1104-16.2016.
- 39. Hernandez, N.S., Ige, K.Y., Mietlicki-Baase, E.G., Molina-Castro, G.C., Turner, C.A., Hayes, M.R., and Schmidt, H.D. (2018). Glucagon-like peptide-1 receptor activation in the ventral tegmental area attenuates cocaine seeking in rats. Neuropsychopharmacology *43*, 2000–2008. https://doi.org/10.1038/s41386-018-0010-3.
- 40. Hernandez, N.S., Weir, V.R., Ragnini, K., Merkel, R., Zhang, Y., Mace, K., Rich, M.T., Christopher Pierce, R., and Schmidt, H.D. (2021). GLP-1 receptor signaling in the laterodorsal tegmental nucleus attenuates cocaine seeking by activating GABAergic circuits that project to the

VTA. Mol. Psychiatr. *26*, 4394–4408. https://doi.org/10.1038/s41380- 020-00957-3.

- 41. Thomsen, M., Holst, J.J., Molander, A., Linnet, K., Ptito, M., and Fink-Jensen, A. (2019). Effects of glucagon-like peptide 1 analogs on alcohol intake in alcohol-preferring vervet monkeys. Psychopharmacology (Berl) *236*, 603–611. https://doi.org/10.1007/s00213-018-5089-z.
- 42. Allen, N.J., and Lyons, D.A. (2018). Glia as architects of central nervous system formation and function. Science *362*, 181–185. https://doi.org/ 10.1126/science.aat0473.
- 43. Nampoothiri, S., Nogueiras, R., Schwaninger, M., and Prevot, V. (2022). Glial cells as integrators of peripheral and central signals in the regulation of energy homeostasis. Nat. Metab. *4*, 813–825. https://doi.org/10.1038/ s42255-022-00610-z.
- 44. Verkhratsky, A., Butt, A., Li, B., Illes, P., Zorec, R., Semyanov, A., Tang, Y., and Sofroniew, M.V. (2023). Astrocytes in human central nervous system diseases: a frontier for new therapies. Signal Transduct. Targeted Ther. *8*, 396. https://doi.org/10.1038/s41392-023-01628-9.
- 45. Timper, K., del Rio-Martin, A., Cremer, A.L., Bremser, S., Alber, J., Gia-valisco, P., Varela, L., Heilinger, C., Nolte, H., Trifunovic, A., et al. (2020). GLP-1 Receptor Signaling in Astrocytes Regulates Fatty Acid Oxidation, Mitochondrial Integrity, and Function. Cell Metabol. *31*, 1189–1205.E13. https://doi.org/10.1016/j.cmet.2020.05.001.
- 46. Zheng, J., Xie, Y., Ren, L., Qi, L., Wu, L., Pan, X., Zhou, J., Chen, Z., and Liu, L. (2021). GLP-1 improves the supportive ability of astrocytes to neurons by promoting aerobic glycolysis in Alzheimer's disease. Mol. Metabol. *47*, 101180. https://doi.org/10.1016/j.molmet.2021.101180.
- 47. Reiner, D.J., Mietlicki-Baase, E.G., McGrath, L.E., Zimmer, D.J., Bence, K.K., Sousa, G.L., Konanur, V.R., Krawczyk, J., Burk, D.H., Kanoski, S.E., et al. (2016). Astrocytes Regulate GLP-1 Receptor-Mediated Effects on Energy Balance. J. Neurosci. *36*, 3531–3540. https://doi.org/10.1523/ JNEUROSCI.3579-15.2016.
- 48. An, J.R., Liu, J.T., Gao, X.M., Wang, Q.F., Sun, G.Y., Su, J.N., Zhang, C., Yu, J.X., Yang, Y.F., and Shi, Y. (2023). Effects of liraglutide on astrocyte polarization and neuroinflammation in db/db mice: focus on iron overload and oxidative stress. Front. Cell. Neurosci. *17*, 1136070. https://doi.org/ 10.3389/fncel.2023.1136070.
- 49. Sterling, J.K., Adetunji, M.O., Guttha, S., Bargoud, A.R., Uyhazi, K.E., Ross, A.G., Dunaief, J.L., and Cui, Q.N. (2020). GLP-1 Receptor Agonist NLY01 Reduces Retinal Inflammation and Neuron Death Secondary to Ocular Hypertension. Cell Rep. *33*, 108271. https://doi.org/10.1016/j.celrep.2020.108271.
- 50. Huang, J., Liu, Y., Cheng, L., Li, J., Zhang, T., Zhao, G., and Zhang, H. (2020). Glucagon-like peptide-1 cleavage product GLP-1(9-36) reduces neuroinflammation from stroke via the activation of insulin-like growth factor 1 receptor in astrocytes. Eur. J. Pharmacol. *887*, 173581. https:// doi.org/10.1016/j.ejphar.2020.173581.
- 51. Vecchiarelli, H.A., and Tremblay, M.È. (2023). Microglial Transcriptional Signatures in the Central Nervous System: Toward A Future of Unraveling Their Function in Health and Disease. Annu. Rev. Genet. *57*, 65–86. https://doi.org/10.1146/annurev-genet-022223-093643.
- 52. Kappe, C., Tracy, L.M., Patrone, C., Iverfeldt, K., and Sjöholm, Å. (2012). GLP-1 secretion by microglial cells and decreased CNS expression in obesity. J. Neuroinflammation *9*, 276. https://doi.org/10.1186/1742- 2094-9-276.
- 53. Qian, Z., Chen, H., Xia, M., Chang, J., Li, X., Ye, S., Wu, S., Jiang, S., Bao, J., Wang, B., et al. (2022). Activation of glucagon-like peptide-1 receptor in microglia attenuates neuroinflammation-induced glial scarring via rescuing Arf and Rho GAP adapter protein 3 expressions after nerve injury. Int. J. Biol. Sci. *18*, 1328–1346. https://doi.org/10.7150/ijbs.68974.
- 54. Yoon, G., Kim, Y.K., and Song, J. (2020). Glucagon-like peptide-1 suppresses neuroinflammation and improves neural structure. Pharmacol. Res. *152*, 104615. https://doi.org/10.1016/j.phrs.2019.104615.
- 55. Jing, F., Zou, Q., Wang, Y., Cai, Z., and Tang, Y. (2021). Activation of microglial GLP-1R in the trigeminal nucleus caudalis suppresses central sensitization of chronic migraine after recurrent nitroglycerin stimulation. J. Headache Pain *22*, 86. https://doi.org/10.1186/s10194-021-01302-x.

Review

- 56. Ventorp, F., Bay-Richter, C., Nagendra, A.S., Janelidze, S., Matsson, V.S., Lipton, J., Nordström, U., Westrin, Å., Brundin, P., and Brundin, L. (2017). Exendin-4 Treatment Improves LPS-Induced Depressive-Like Behavior Without Affecting Pro-Inflammatory Cytokines. J. Parkinsons Dis. *7*, 263–273. https://doi.org/10.3233/JPD-171068.
- 57. Yun, S.P., Kam, T.I., Panicker, N., Kim, S., Oh, Y., Park, J.S., Kwon, S.H., Park, Y.J., Karuppagounder, S.S., Park, H., et al. (2018). Block of A1 astrocyte conversion by microglia is neuroprotective in models of Parkinson's disease. Nat. Med. *24*, 931–938. https://doi.org/10.1038/s41591- 018-0051-5.
- 58. McGarry, A., Rosanbalm, S., Leinonen, M., Olanow, C.W., To, D., Bell, A., Lee, D., Chang, J., Dubow, J., Dhall, R., et al. (2024). Safety, tolerability, and efficacy of NLY01 in early untreated Parkinson's disease: a randomised, double-blind, placebo-controlled trial. Lancet Neurol. *23*, 37–45. https://doi.org/10.1016/S1474-4422(23)00378-2.
- 59. Nave, K.A., and Werner, H.B. (2014). Myelination of the nervous system: mechanisms and functions. Annu. Rev. Cell Dev. Biol. *30*, 503–533. https://doi.org/10.1146/annurev-cellbio-100913-013101.
- 60. Smith, C., Patterson-Cross, R., Woodward, O., Lewis, J., Chiarugi, D., Merkle, F., Gribble, F., Reimann, F., and Adriaenssens, A. (2022). A comparative transcriptomic analysis of glucagon-like peptide-1 receptor- and glucose-dependent insulinotropic polypeptide receptor-ex-pressing cells in the hypothalamus. Appetite *174*, 106022. https://doi. org/10.1016/j.appet.2022.106022.
- 61. Gharagozloo, M., Galleguillos, D., Jank, L., Sotirchos, E.S., Smith, M.D., Garton, T., Kumar, S., Hussein, O., Potluri, S., Taylor, M., et al. (2023). The Effects of NLY01, a Novel Glucagon-Like Peptide-1 Receptor Agonist, on Cuprizone-Induced Demyelination and Remyelination: Challenges and Future Perspectives. Neurotherapeutics *20*, 1229–1240. https://doi. org/10.1007/s13311-023-01390-4.
- 62. Yang, A.C., Vest, R.T., Kern, F., Lee, D.P., Agam, M., Maat, C.A., Losada, P.M., Chen, M.B., Schaum, N., Khoury, N., et al. (2022). A human brain vascular atlas reveals diverse mediators of Alzheimer's risk. Nature *603*, 885–892. https://doi.org/10.1038/s41586-021-04369-3.
- 63. Dong, M., Wen, S., and Zhou, L. (2022). The Relationship Between the Blood-Brain-Barrier and the Central Effects of Glucagon-Like Peptide-1 Receptor Agonists and Sodium-Glucose Cotransporter-2 Inhibitors. Diabetes Metab. Syndr. Obes. *15*, 2583–2597. https://doi.org/10.2147/ DMSO.S375559.
- 64. Fu, Z., Gong, L., Liu, J., Wu, J., Barrett, E.J., Aylor, K.W., and Liu, Z. (2020). Brain Endothelial Cells Regulate Glucagon-Like Peptide 1 Entry Into the Brain via a Receptor-Mediated Process. Front. Physiol. *11*, 555. https://doi.org/10.3389/fphys.2020.00555.
- 65. Fukuda, S., Nakagawa, S., Tatsumi, R., Morofuji, Y., Takeshita, T., Hayashi, K., Tanaka, K., Matsuo, T., and Niwa, M. (2016). Glucagon-Like Peptide-1 Strengthens the Barrier Integrity in Primary Cultures of Rat Brain Endothelial Cells Under Basal and Hyperglycemia Conditions. J. Mol. Neurosci. *59*, 211–219. https://doi.org/10.1007/s12031-015- 0696-1.
- 66. Xie, Z., Enkhjargal, B., Nathanael, M., Wu, L., Zhu, Q., Zhang, T., Tang, J., and Zhang, J.H. (2021). Exendin-4 Preserves Blood-Brain Barrier Integrity *via* Glucagon-Like Peptide 1 Receptor/Activated Protein Kinase-Dependent Nuclear Factor-Kappa B/Matrix Metalloproteinase-9 Inhibition After Subarachnoid Hemorrhage in Rat. Front. Mol. Neurosci. *14*, 750726. https://doi.org/10.3389/fnmol.2021.750726.
- 67. Li, Y., Perry, T., Kindy, M.S., Harvey, B.K., Tweedie, D., Holloway, H.W., Powers, K., Shen, H., Egan, J.M., Sambamurti, K., et al. (2009). GLP-1 receptor stimulation preserves primary cortical and dopaminergic neurons in cellular and rodent models of stroke and Parkinsonism. Proc. Natl. Acad. Sci. USA *106*, 1285–1290. https://doi.org/10.1073/pnas. 0806720106.
- 68. He, X., Wen, S., Tang, X., Wen, Z., Zhang, R., Li, S., Gao, R., Wang, J., Zhu, Y., Fang, D., et al. (2024). Glucagon-like peptide-1 receptor agonists rescued diabetic vascular endothelial damage through suppression of aberrant STING signaling. Acta Pharm. Sin. B *14*, 2613–2630. https:// doi.org/10.1016/j.apsb.2024.03.011.
- 69. Siekmann, A.F. (2023). Biology of vascular mural cells. Development *150*, dev200271. https://doi.org/10.1242/dev.200271.
- 70. Grutzendler, J., and Nedergaard, M. (2019). Cellular Control of Brain Capillary Blood Flow: In Vivo Imaging Veritas. Trends Neurosci. *42*, 528–536. https://doi.org/10.1016/j.tins.2019.05.009.
- 71. Nizari, S., Basalay, M., Chapman, P., Korte, N., Korsak, A., Christie, I.N., Theparambil, S.M., Davidson, S.M., Reimann, F., Trapp, S., et al. (2021). Glucagon-like peptide-1 (GLP-1) receptor activation dilates cerebral arterioles, increases cerebral blood flow, and mediates remote (pre)conditioning neuroprotection against ischaemic stroke. Basic Res. Cardiol. *116*, 32. https://doi.org/10.1007/s00395-021-00873-9.
- 72. Bailey, J., Coucha, M., Bolduc, D.R., Burnett, F.N., Barrett, A.C., Ghaly, M., and Abdelsaid, M. (2022). GLP-1 receptor nitration contributes to loss of brain pericyte function in a mouse model of diabetes. Diabetologia *65*, 1541–1554. https://doi.org/10.1007/s00125-022-05730-5.
- 73. Oezer, K., Kolibabka, M., Gassenhuber, J., Dietrich, N., Fleming, T., Schlotterer, A., Morcos, M., Wohlfart, P., and Hammes, H.P. (2023). The effect of GLP-1 receptor agonist lixisenatide on experimental diabetic retinopathy. Acta Diabetol. *60*, 1551–1565. https://doi.org/10. 1007/s00592-023-02135-7.
- 74. Lin, W.J., Ma, X.F., Hao, M., Zhou, H.R., Yu, X.Y., Shao, N., Gao, X.Y., and Kuang, H.Y. (2018). Liraglutide attenuates the migration of retinal pericytes induced by advanced glycation end products. Peptides *105*, 7–13. https://doi.org/10.1016/j.peptides.2018.05.003.
- 75. Zimmerman, B., Kundu, P., Rooney, W.D., and Raber, J. (2021). The Effect of High Fat Diet on Cerebrovascular Health and Pathology: A Species Comparative Review. Molecules *26*, 3406. https://doi.org/10. 3390/molecules26113406.
- 76. Yamamoto, M., Guo, D.H., Hernandez, C.M., and Stranahan, A.M. (2019). Endothelial Adora2a Activation Promotes Blood-Brain Barrier Breakdown and Cognitive Impairment in Mice with Diet-Induced Insulin Resistance. J. Neurosci. *39*, 4179–4192. https://doi.org/10.1523/ JNEUROSCI.2506-18.2019.
- 77. Chen, B., de Launoit, E., Meseguer, D., Garcia Caceres, C., Eichmann, A., Renier, N., and Schneeberger, M. (2024). The interactions between energy homeostasis and neurovascular plasticity. Nat. Rev. Endocrinol. *x*, x. https://doi.org/10.1038/s41574-024-01021-8.
- 78. Chen, B., de Launoit, E., Renier, N., and Schneeberger, M. (2024). Maternal nutritional programming shapes the cerebral landscape. Trends Endocrinol. Metabol. *35*, 367–370. https://doi.org/10.1016/j.tem.2023. 10.008.
- 79. Amen, D.G., Wu, J., George, N., and Newberg, A. (2020). Patterns of Regional Cerebral Blood Flow as a Function of Obesity in Adults. J. Alzheimers Dis. *77*, 1331–1337. https://doi.org/10.3233/JAD-200655.
- 80. Carter, K.J., Ward, A.T., Kellawan, J.M., Harrell, J.W., Peltonen, G.L., Roberts, G.S., Al-Subu, A., Hagen, S.A., Serlin, R.C., Eldridge, M.W., et al. (2023). Reduced basal macrovascular and microvascular cerebral blood flow in young adults with metabolic syndrome: potential mechanisms. J. Appl. Physiol. *135*, 94–108. https://doi.org/10.1152/japplphysiol.00688.
- 81. Pasha, E.P., Birdsill, A.C., Oleson, S., Haley, A.P., and Tanaka, H. (2017). Impacts of Metabolic Syndrome Scores on Cerebrovascular Conductance Are Mediated by Arterial Stiffening. Am. J. Hypertens. *31*, 72–79. https://doi.org/10.1093/ajh/hpx132.
- 82. Birdsill, A.C., Carlsson, C.M., Willette, A.A., Okonkwo, O.C., Johnson, S.C., Xu, G., Oh, J.M., Gallagher, C.L., Koscik, R.L., Jonaitis, E.M., et al. (2013). Low cerebral blood flow is associated with lower memory function in metabolic syndrome. Obesity *21*, 1313–1320. https://doi. org/10.1002/oby.20170.
- 83. Alwatban, M.R., Aaron, S.E., Kaufman, C.S., Barnes, J.N., Brassard, P., Ward, J.L., Miller, K.B., Howery, A.J., Labrecque, L., and Billinger, S.A. (2021). Effects of age and sex on middle cerebral artery blood velocity and flow pulsatility index across the adult lifespan. J. Appl. Physiol. *130*, 1675–1683. https://doi.org/10.1152/japplphysiol.00926.2020.
- 84. Janelidze, S., Hertze, J., Nägga, K., Nilsson, K., Nilsson, C., Swedish BioFINDER Study Group, Wennström, M., van Westen, D., Blennow, K., Zetterberg, H., and Hansson, O. (2017). Increased blood-brain barrier permeability is associated with dementia and diabetes but not amyloid pathology or APOE genotype. Neurobiol. Aging *51*, 104–112. https:// doi.org/10.1016/j.neurobiolaging.2016.11.017.

d CellPress

- 85. van Sloten, T.T., Sedaghat, S., Carnethon, M.R., Launer, L.J., and Stehouwer, C.D.A. (2020). Cerebral microvascular complications of type 2 diabetes: stroke, cognitive dysfunction, and depression. Lancet Diabetes Endocrinol. *8*, 325–336. https://doi.org/10.1016/S2213-8587(19) 30405-X.
- 86. Huber, G., Ogrodnik, M., Wenzel, J., Stölting, I., Huber, L., Will, O., Peschke, E., Matschl, U., Hövener, J.B., Schwaninger, M., et al. (2021). Telmisartan prevents high-fat diet-induced neurovascular impairments and reduces anxiety-like behavior. J. Cerebr. Blood Flow Metabol. *41*, 2356–2369. https://doi.org/10.1177/0271678X211003497.
- 87. Boitard, C., Etchamendy, N., Sauvant, J., Aubert, A., Tronel, S., Marighetto, A., Layé, S., and Ferreira, G. (2012). Juvenile, but not adult
exposure to high-fat diet impairs relational memory and hippocampal neurogenesis in mice. Hippocampus *22*, 2095–2100. https://doi.org/10. 1002/hipo.22032.
- 88. Pannacciulli, N., Le, D.S.N.T., Salbe, A.D., Chen, K., Reiman, E.M., Tataranni, P.A., and Krakoff, J. (2007). Postprandial glucagon-like peptide-1 (GLP-1) response is positively associated with changes in neuronal activity of brain areas implicated in satiety and food intake regulation in hu-mans. Neuroimage *35*, 511–517. https://doi.org/10.1016/j.neuroimage. 2006.12.035.
- 89. Lin, M.H., Cheng, P.C., Hsiao, P.J., Chen, S.C., Hung, C.H., Kuo, C.H., Huang, S.K., and Clair Chiou, H.Y. (2023). The GLP-1 receptor agonist exenatide ameliorates neuroinflammation, locomotor activity, and anxiety-like behavior in mice with diet-induced obesity through the modulation of microglial M2 polarization and downregulation of SR-A4. Int. Immunopharm. *115*, 109653. https://doi.org/10.1016/j.intimp.2022. 109653.
- 90. Bułdak, Ł., Machnik, G., Skudrzyk, E., Boldys, A., and Okopien, B. (2019). The impact of exenatide (a GLP-1 agonist) on markers of inflammation and oxidative stress in normal human astrocytes subjected to various glycemic conditions. Exp. Ther. Med. *17*, 2861–2869. https:// doi.org/10.3892/etm.2019.7245.
- 91. Zanotto, C., Simão, F., Gasparin, M.S., Biasibetti, R., Tortorelli, L.S., Nardin, P., and Gonçalves, C.A. (2017). Exendin-4 Reverses Biochemical and Functional Alterations in the Blood-Brain and Blood-CSF Barriers in Diabetic Rats. Mol. Neurobiol. *54*, 2154–2166. https://doi.org/10. 1007/s12035-016-9798-1.
- 92. Li, Q., Jia, M., Yan, Z., Li, Q., Sun, F., He, C., Li, Y., Zhou, X., Zhang, H., Liu, X., et al. (2021). Activation of Glucagon-Like Peptide-1 Receptor Ameliorates Cognitive Decline in Type 2 Diabetes Mellitus Through a Metabolism-Independent Pathway. J. Am. Heart Assoc. *10*, e020734. https://doi.org/10.1161/JAHA.120.020734.
- 93. Marques-Neto, S.R., Castiglione, R.C., Pontes, A., Oliveira, D.F., Ferraz, E.B., Nascimento, J.H.M., and Bouskela, E. (2016). Effects of Incretin-Based Therapies on Neuro-Cardiovascular Dynamic Changes Induced by High Fat Diet in Rats. PLoS One *11*, e0148402. https://doi.org/10. 1371/journal.pone.0148402.
- 94. Baylan, U., Korn, A., Emmens, R.W., Schalkwijk, C.G., Niessen, H.W.M., Krijnen, P.A.J., and Simsek, S. (2022). Liraglutide treatment attenuates inflammation markers in the cardiac, cerebral and renal microvasculature in streptozotocin-induced diabetic rats. Eur. J. Clin. Invest. *52*, e13807. https://doi.org/10.1111/eci.13807.
- 95. Carranza-Naval, M.J., Del Marco, A., Hierro-Bujalance, C., Alves-Martinez, P., Infante-Garcia, C., Vargas-Soria, M., Herrera, M., Barba-Cordoba, B., Atienza-Navarro, I., Lubian-Lopez, S., and Garcia-Alloza, M. (2021). Liraglutide Reduces Vascular Damage, Neuronal Loss, and Cognitive Impairment in a Mixed Murine Model of Alzheimer's Disease and Type 2 Diabetes. Front. Aging Neurosci. *13*, 741923. https://doi. org/10.3389/fnagi.2021.741923.
- 96. Hardigan, T., Abdul, Y., and Ergul, A. (2016). Linagliptin reduces effects of ET-1 and TLR2-mediated cerebrovascular hyperreactivity in diabetes. Life Sci. *159*, 90–96. https://doi.org/10.1016/j.lfs.2016.02.067.
- 97. Abdelsaid, M., Williams, R., Hardigan, T., and Ergul, A. (2016). Linagliptin attenuates diabetes-induced cerebral pathological neovascularization in a blood glucose-independent manner: Potential role of ET-1. Life Sci. *159*, 83–89. https://doi.org/10.1016/j.lfs.2015.11.026.
- 98. Elabi, O.F., Karampatsi, D., Vercalsteren, E., Lietzau, G., Nyström, T., Klein, T., Darsalia, V., Patrone, C., and Paul, G. (2023). DPP-4 Inhibitor

Cell Metabolism Review

and Sulfonylurea Differentially Reverse Type 2 Diabetes-Induced Blood-Brain Barrier Leakage and Normalize Capillary Pericyte Coverage. Diabetes *72*, 405–414. https://doi.org/10.2337/db22-0674.

- 99. Hardigan, T., Yasir, A., Abdelsaid, M., Coucha, M., El-Shaffey, S., Li, W., Johnson, M.H., and Ergul, A. (2016). Linagliptin treatment improves cerebrovascular function and remodeling and restores reduced cerebral perfusion in Type 2 diabetes. Am. J. Physiol. Regul. Integr. Comp. Physiol. *311*, R466–R477. https://doi.org/10.1152/ajpregu.00057.2016.
- 100. Fuchs, H., Binder, R., and Greischel, A. (2009). Tissue distribution of the novel DPP-4 inhibitor BI 1356 is dominated by saturable binding to its target in rats. Biopharm. Drug Dispos. *30*, 229–240. https://doi.org/10. 1002/bdd.662.
- 101. Darsalia, V., Larsson, M., Lietzau, G., Nathanson, D., Nyström, T., Klein,
T., and Patrone, C. (2016). Gliptin-mediated neuroprotection against stroke requires chronic pretreatment and is independent of glucagon-like peptide-1 receptor. Diabetes Obes. Metabol. *18*, 537–541. https:// doi.org/10.1111/dom.12641.
- 102. Ölmestig, J., Marlet, I.R., Vilsboll, T., Rungby, J., Rostrup, E., Lambertsen, K.L., and Kruuse, C. (2022). A single dose of exenatide had no effect on blood flow velocity in the middle cerebral artery in elderly healthy volunteers: Randomized, placebo-controlled, double-blind clinical trial. Front. Aging Neurosci. *14*, 899389. https://doi.org/10.3389/fnagi.2022. 899389.
- 103. Lin, L., Wang, X., and Yu, Z. (2016). Ischemia-reperfusion Injury in the Brain: Mechanisms and Potential Therapeutic Strategies. Biochem. Pharmacol. *5*, 213. https://doi.org/10.4172/2167-0501.1000213.
- 104. Quiñones-Ossa, G.A., Lobo, C., Garcia-Ballestas, E., Florez, W.A., Moscote-Salazar, L.R., and Agrawal, A. (2021). Obesity and Stroke: Does the Paradox Apply for Stroke? Neurointervention *16*, 9–19. https://doi.org/ 10.5469/neuroint.2020.00108.
- 105. Chen, B., and Schneeberger, M. (2024). Neuro-Adipokine Crosstalk in Alzheimer's Disease. Int. J. Mol. Sci. *25*, 5932. https://doi.org/10.3390/ iims25115932.
- 106. Obadia, N., Lessa, M.A., Daliry, A., Silvares, R.R., Gomes, F., Tibiriçá, E., and Estato, V. (2017). Cerebral microvascular dysfunction in metabolic syndrome is exacerbated by ischemia-reperfusion injury. BMC Neurosci. *18*, 67. https://doi.org/10.1186/s12868-017-0384-x.
- 107. Li, W., Prakash, R., Chawla, D., Du, W., Didion, S.P., Filosa, J.A., Zhang, Q., Brann, D.W., Lima, V.V., Tostes, R.C., and Ergul, A. (2013). Early effects of high-fat diet on neurovascular function and focal ischemic brain injury. Am. J. Physiol. Regul. Integr. Comp. Physiol. *304*, R1001–R1008. https://doi.org/10.1152/ajpregu.00523.2012.
- 108. Kuroki, T., Tanaka, R., Shimada, Y., Yamashiro, K., Ueno, Y., Shimura, H., Urabe, T., and Hattori, N. (2016). Exendin-4 Inhibits Matrix Metalloproteinase-9 Activation and Reduces Infarct Growth After Focal Cerebral Ischemia in Hyperglycemic Mice. Stroke *47*, 1328–1335. https://doi.org/10.1161/STROKEAHA.116.012934.
- 109. Deng, J., Zhang, J., Feng, C., Xiong, L., and Zuo, Z. (2014). Critical role of matrix metalloprotease-9 in chronic high fat diet-induced cerebral vascular remodelling and increase of ischaemic brain injury in mice. Cardiovasc. Res. *103*, 473–484. https://doi.org/10.1093/cvr/cvu154.
- 110. Zhang, Z., Yan, J., and Shi, H. (2016). Role of Hypoxia Inducible Factor 1 in Hyperglycemia-Exacerbated Blood-Brain Barrier Disruption in Ischemic Stroke. Neurobiol. Dis. *95*, 82–92. https://doi.org/10.1016/j. nbd.2016.07.012.
- 111. Soejima, Y., Hu, Q., Krafft, P.R., Fujii, M., Tang, J., and Zhang, J.H. (2013). Hyperbaric oxygen preconditioning attenuates hyperglycemiaenhanced hemorrhagic transformation by inhibiting matrix metalloproteinases in focal cerebral ischemia in rats. Exp. Neurol. *247*, 737–743. https://doi.org/10.1016/j.expneurol.2013.03.019.
- 112. Karampatsi, D., Zabala, A., Wilhelmsson, U., Dekens, D., Vercalsteren, E., Larsson, M., Nyström, T., Pekny, M., Patrone, C., and Darsalia, V. (2021). Diet-induced weight loss in obese/diabetic mice normalizes glucose metabolism and promotes functional recovery after stroke. Cardiovasc. Diabetol. *20*, 240. https://doi.org/10.1186/s12933-021- 01426-z.

Review

- 113. Yin, Q., Ma, J., Han, X., Zhang, H., Wang, F., Zhuang, P., and Zhang, Y. (2021). Spatiotemporal variations of vascular endothelial growth factor in the brain of diabetic cognitive impairment. Pharmacol. Res. *163*, 105234. https://doi.org/10.1016/j.phrs.2020.105234.
- 114. Tsao, C.W., Aday, A.W., Almarzooq, Z.I., Anderson, C.A.M., Arora, P., Avery, C.L., Baker-Smith, C.M., Beaton, A.Z., Boehme, A.K., Buxton, A.E., et al. (2023). Heart Disease and Stroke Statistics-2023 Update: A Report From the American Heart Association. Circulation *147*, e93– e621. https://doi.org/10.1161/CIR.0000000000001123.
- 115. Lansberg, M.G., Schrooten, M., Bluhmki, E., Thijs, V.N., and Saver, J.L. (2009). Treatment time-specific number needed to treat estimates for tissue plasminogen activator therapy in acute stroke based on shifts over the entire range of the modified Rankin Scale. Stroke *40*, 2079–2084. https://doi.org/10.1161/STROKEAHA.108.540708.
- 116. Prabhakaran, S., Ruff, I., and Bernstein, R.A. (2015). Acute stroke intervention: a systematic review. JAMA *313*, 1451–1462. https://doi.org/ 10.1001/jama.2015.3058.
- 117. Lee, C.H., Yan, B., Yoo, K.Y., Choi, J.H., Kwon, S.H., Her, S., Sohn, Y., Hwang, I.K., Cho, J.H., Kim, Y.M., and Won, M.H. (2011). Ischemiainduced changes in glucagon-like peptide-1 receptor and neuroprotective effect of its agonist, exendin-4, in experimental transient cerebral ischemia. J. Neurosci. Res. *89*, 1103–1113. https://doi.org/10.1002/ jnr.22596.
- 118. Zan, L., Zhang, X., Xi, Y., Wu, H., Song, Y., Teng, G., Li, H., Qi, J., and Wang, J. (2014). Src regulates angiogenic factors and vascular permeability after focal cerebral ischemia-reperfusion. Neuroscience *262*, 118–128. https://doi.org/10.1016/j.neuroscience.2013.12.060.
- 119. Kim, S., Jeong, J., Jung, H.S., Kim, B., Kim, Y.E., Lim, D.S., Kim, S.D., and Song, Y.S. (2017). Anti-inflammatory Effect of Glucagon Like Peptide-1 Receptor Agonist, Exendin-4, through Modulation of IB1/ JIP1 Expression and JNK Signaling in Stroke. Exp. Neurobiol. *26*, 227–239. https://doi.org/10.5607/en.2017.26.4.227.
- 120. Liu, C., Sun, S., Xie, J., Li, H., Li, T., Wu, Q., Zhang, Y., Bai, X., Wang, J., Wang, X., et al. (2022). GLP-1R Agonist Exendin-4 Protects Against Hemorrhagic Transformation Induced by rtPA After Ischemic Stroke via the Wnt/b-Catenin Signaling Pathway. Mol. Neurobiol. *59*, 3649–3664. https://doi.org/10.1007/s12035-022-02811-9.
- 121. Kasprzak, A. (2020). Angiogenesis-Related Functions of Wnt Signaling in Colorectal Carcinogenesis. Cancers *12*, 3601. https://doi.org/10.3390/ cancers12123601.
- 122. Ozkul-Wermester, O., Guegan-Massardier, E., Triquenot, A., Borden, A., Perot, G., and Gérardin, E. (2014). Increased blood-brain barrier permeability on perfusion computed tomography predicts hemorrhagic transformation in acute ischemic stroke. Eur. Neurol. *72*, 45–53. https://doi. org/10.1159/000358297.
- 123. Shan, Y., Tan, S., Lin, Y., Liao, S., Zhang, B., Chen, X., Wang, J., Deng, Z., Zeng, Q., Zhang, L., et al. (2019). The glucagon-like peptide-1 receptor agonist reduces inflammation and blood-brain barrier breakdown in an astrocyte-dependent manner in experimental stroke. J. Neuroinflammation *16*, 242. https://doi.org/10.1186/s12974-019-1638-6.
- 124. Augestad, I.L., Dekens, D., Karampatsi, D., Elabi, O., Zabala, A., Pintana, H., Larsson, M., Nyström, T., Paul, G., Darsalia, V., and Patrone, C. (2022). Normalisation of glucose metabolism by exendin-4 in the chronic phase after stroke promotes functional recovery in male diabetic mice. Br. J. Pharmacol. 179, 677-694. https://doi.org/10.1111/bph.155.
- 125. Gonçalves, A., Lin, C.M., Muthusamy, A., Fontes-Ribeiro, C., Ambrosio, A.F., Abcouwer, S.F., Fernandes, R., and Antonetti, D.A. (2016). Protec-tive Effect of a GLP-1 Analog on Ischemia-Reperfusion Induced Blood-Retinal Barrier Breakdown and Inflammation. Invest. Ophthalmol. Vis. Sci. *57*, 2584–2592. https://doi.org/10.1167/iovs.15-19006.
- 126. Chien, C.T., Jou, M.J., Cheng, T.Y., Yang, C.H., Yu, T.Y., and Li, P.C. (2015). Exendin-4-loaded PLGA microspheres relieve cerebral ischemia/reperfusion injury and neurologic deficits through long-lasting bioactivity-mediated phosphorylated Akt/eNOS signaling in rats. J. Cerebr. Blood Flow Metabol. *35*, 1790–1803. https://doi.org/10. 1038/jcbfm.2015.126.
- 127. Basalay, M.V., Davidson, S.M., and Yellon, D.M. (2019). Neuroprotection in Rats Following Ischaemia-Reperfusion Injury by GLP-1 Analogues-

Liraglutide and Semaglutide. Cardiovasc. Drugs Ther. *33*, 661–667. https://doi.org/10.1007/s10557-019-06915-8.

- 128. Li, Y., and Gong, M. (2021). Analysis of the neuroprotective effect of GLP-1 receptor agonist peptide on cerebral ischemia-reperfusion injury by Quantitative Proteomics Mass Spectrometry. Brain Behav. *11*, e02190. https://doi.org/10.1002/brb3.2190.
- 129. Yang, L., Cheng, J., Shi, G., Zhang, C., Du, Y., Chen, L., Qiao, H., Chen, R., and Zhang, X. (2022). Liraglutide Ameliorates Cerebral Ischemia in Mice via Antipyroptotic Pathways. Neurochem. Res. *47*, 1904–1916. https://doi.org/10.1007/s11064-022-03574-4.
- 130. Tu, X.K., Chen, P.P., Chen, J.Y., Ding, Y.H., Chen, Q., and Shi, S.S. (2023). GLP-1R knockdown abrogates the protective effects of liraglutide on ischaemic stroke via inhibition of M2 polarisation and activation of NLRP3 inflammasome by reducing Nrf2 activation. Neuropharmacology *237*, 109603. https://doi.org/10.1016/j.neuropharm.2023.109603.
- 131. Ward, R., Li, W., Abdul, Y., Jackson, L., Dong, G., Jamil, S., Filosa, J., Fagan, S.C., and Ergul, A. (2019). NLRP3 inflammasome inhibition with MCC950 improves diabetes-mediated cognitive impairment and vasoneuronal remodeling after ischemia. Pharmacol. Res. *142*, 237–250. https://doi.org/10.1016/j.phrs.2019.01.035.
- 132. Chen, X., Huang, Q., Feng, J., Xiao, Z., Zhang, X., and Zhao, L. (2021). GLP-1 alleviates NLRP3 inflammasome-dependent inflammation in perivascular adipose tissue by inhibiting the NF-kB signalling pathway. J. Int. Med. Res. *49*, 300060521992981. https://doi.org/10.1177/0300060521992981.
- 133. Jin, H., Zhu, Y., Li, Y., Ding, X., Ma, W., Han, X., and Wang, B. (2019). BDNF-mediated mitophagy alleviates high-glucose-induced brain microvascular endothelial cell injury. Apoptosis *24*, 511–528. https:// doi.org/10.1007/s10495-019-01535-x.
- 134. Liu, Y., Hu, Z., Wang, J., Liao, Y., and Shu, L. (2023). Puerarin alleviates depressive-like behaviors in high-fat diet-induced diabetic mice via modulating hippocampal GLP-1R/BDNF/TrkB signaling. Nutr. Neurosci. *26*, 997–1010. https://doi.org/10.1080/1028415X.2022.2112439.
- 135. Dong, Q., Teng, S.W., Wang, Y., Qin, F., Li, Y., Ai, L.L., and Yu, H. (2019). Sitagliptin protects the cognition function of the Alzheimer's disease mice through activating glucagon-like peptide-1 and BDNF-TrkB signalings. Neurosci. Lett. *696*, 184–190. https://doi.org/10.1016/j.neulet. 2018.12.041.
- 136. Kutlu, M.D., Kose, S., and Akillioglu, K. (2023). GLP-1 agonist Liraglutide prevents MK-801-induced schizophrenia-like behaviors and BDNF, CREB, p-CREB, Trk-B expressions in the hippocampus and prefrontal cortex in Balb/c mice. Behav. Brain Res. *445*, 114386. https://doi.org/ 10.1016/j.bbr.2023.114386.
- 137. Deng, C., Cao, J., Han, J., Li, J., Li, Z., Shi, N., and He, J. (2018). Liraglutide Activates the Nrf2/HO-1 Antioxidant Pathway and Protects Brain Nerve Cells against Cerebral Ischemia in Diabetic Rats. Comput. Intell. Neurosci. *2018*, 3094504. https://doi.org/10.1155/2018/3094504.
- 138. Briyal, S., Shah, S., and Gulati, A. (2014). Neuroprotective and antiapoptotic effects of liraglutide in the rat brain following focal cerebral ischemia. Neuroscience *281*, 269–281. https://doi.org/10.1016/j.neuroscience.2014.09.064.
- 139. Sato, K., Kameda, M., Yasuhara, T., Agari, T., Baba, T., Wang, F., Shinko, A., Wakamori, T., Toyoshima, A., Takeuchi, H., et al. (2013). Neuroprotective effects of liraglutide for stroke model of rats. Int. J. Mol. Sci. *14*, 21513–21524. https://doi.org/10.3390/ijms141121513.
- 140. Chen, Y., Zhang, X., He, J., Xie, Y., and Yang, Y. (2018). Delayed Administration of the Glucagon-Like Peptide 1 Analog Liraglutide Promoting Angiogenesis after Focal Cerebral Ischemia in Mice. J. Stroke Cerebrovasc. Dis. *27*, 1318–1325. https://doi.org/10.1016/j.jstrokecerebrovasdis.2017.12.015.
- 141. Dong, W., Miao, Y., Chen, A., Cheng, M., Ye, X., Song, F., and Zheng, G. (2017). Delayed administration of the GLP-1 receptor agonist liraglutide improves metabolic and functional recovery after cerebral ischemia in rats. Neurosci. Lett. *641*, 1–7. https://doi.org/10.1016/j.neulet.2017. 01.045.
- 142. Li, P.C., Liu, L.F., Jou, M.J., and Wang, H.K. (2016). The GLP-1 receptor agonists exendin-4 and liraglutide alleviate oxidative stress and cognitive and micturition deficits induced by middle cerebral artery occlusion in

diabetic mice. BMC Neurosci. *17*, 37. https://doi.org/10.1186/s12868- 016-0272-9.

- 143. Zhang, Q., Liu, C., Shi, R., Zhou, S., Shan, H., Deng, L., Chen, T., Guo, Y., Zhang, Z., Yang, G.Y., et al. (2022). Blocking C3d⁺ /GFAP⁺ A1 Astrocyte Conversion with Semaglutide Attenuates Blood-Brain Barrier Disruption in Mice after Ischemic Stroke. Aging Dis. *13*, 943–959. https://doi.org/10. 14336/AD.2021.1029.
- 144. Yang, X., Feng, P., Zhang, X., Li, D., Wang, R., Ji, C., Li, G., and Hölscher, C. (2019). The diabetes drug semaglutide reduces infarct size, inflammation, and apoptosis, and normalizes neurogenesis in a rat model of stroke. Neuropharmacology *158*, 107748. https://doi.org/10.1016/j.neuropharm.2019.107748.
- 145. Abdel-Latif, R.G., Heeba, G.H., Taye, A., and Khalifa, M.M.A. (2018). Lix-isenatide ameliorates cerebral ischemia-reperfusion injury via GLP-1 receptor dependent/independent pathways. Eur. J. Pharmacol. *833*, 145–154. https://doi.org/10.1016/j.ejphar.2018.05.045.
- 146. Abdel-Latif, R.G., Heeba, G.H., Taye, A., and Khalifa, M.M.A. (2018). Lixisenatide, a novel GLP-1 analog, protects against cerebral ischemia/reperfusion injury in diabetic rats. Naunyn-Schmiedeberg's Arch. Pharma-col. *391*, 705–717. https://doi.org/10.1007/s00210-018-1497-1.
- 147. Gad, S.N., Nofal, S., Raafat, E.M., and Ahmed, A.A.E. (2020). Lixisenatide Reduced Damage in Hippocampus CA1 Neurons in a Rat Model of Cerebral Ischemia-Reperfusion Possibly Via the ERK/P38 Signaling Pathway. J. Mol. Neurosci. *70*, 1026–1037. https://doi.org/10.1007/ s12031-020-01497-9.
- 148. Darsalia, V., Ortsäter, H., Olverling, A., Darlöf, E., Wolbert, P., Nyström, T., Klein, T., Sjöholm, Å., and Patrone, C. (2013). The DPP-4 inhibitor linagliptin counteracts stroke in the normal and diabetic mouse brain: a comparison with glimepiride. Diabetes *62*, 1289–1296. https://doi.org/10. 2337/db12-0988.
- 149. Ma, M., Hasegawa, Y., Koibuchi, N., Toyama, K., Uekawa, K., Nakagawa, T., Lin, B., and Kim-Mitsuyama, S. (2015). DPP-4 inhibition with linagliptin ameliorates cognitive impairment and brain atrophy induced by transient cerebral ischemia in type 2 diabetic mice. Cardiovasc. Diabetol. *14*, 54. https://doi.org/10.1186/s12933-015-0218-z.
- 150. Chiazza, F., Tammen, H., Pintana, H., Lietzau, G., Collino, M., Nyström,
T., Klein, T., Darsalia, V., and Patrone, C. (2018). The effect of DPP-4 inhibition to improve functional outcome after stroke is mediated by the SDF-1a/CXCR4 pathway. Cardiovasc. Diabetol. *17*, 60. https://doi.org/ 10.1186/s12933-018-0702-3.
- 151. Sadri, F., Rezaei, Z., and Fereidouni, M. (2022). The significance of the SDF-1/CXCR4 signaling pathway in the normal development. Mol. Biol. Rep. *49*, 3307–3320. https://doi.org/10.1007/s11033-021-07069-3.
- 152. Mi, D.H., Fang, H.J., Zheng, G.H., Liang, X.H., Ding, Y.R., Liu, X., and Liu, L.P. (2019). DPP-4 inhibitors promote proliferation and migration of rat brain microvascular endothelial cells under hypoxic/high-glucose conditions, potentially through the SIRT1/HIF-1/VEGF pathway. CNS Neurosci. Ther. *25*, 323–332. https://doi.org/10.1111/cns.13042.
- 153. Al-Awar, A., Almási, N., Szabo, R., Takacs, I., Murlasits, Z., Szucs, G.,
Torok, S., Posa, A., Varga, C., and Kupai, K. (2018). Novel Potentials of the DPP-4 Inhibitor Sitagliptin against Ischemia-Reperfusion (I/R) Injury in Rat Ex-Vivo Heart Model. Int. J. Mol. Sci. *19*, 3226. https://doi.org/ 10.3390/ijms19103226.
- 154. Xiong, J., Wang, Z., Bai, J., Cheng, K., Liu, Q., and Ni, J. (2023). Calcitonin gene-related peptide: a potential protective agent in cerebral ischemiareperfusion injury. Front. Neurosci. *17*, 1184766. https://doi.org/10. 3389/fnins.2023.1184766.
- 155. Guo, Y., Zhang, Q., Chen, H., Jiang, Y., and Gong, P. (2019). Overexpression of calcitonin gene-related peptide protects mouse cerebral microvascular endothelial cells from high-glucose-induced damage via ERK/ HIF-1/VEGF signaling. J. Physiol. Sci. *69*, 939–952. https://doi.org/10. 1007/s12576-019-00708-2.
- 156. Sorby-Adams, A.J., Vink, R., and Turner, R.J. (2018). Large animal models of stroke and traumatic brain injury as translational tools. Am. J. Physiol. Regul. Integr. Comp. Physiol. *315*, R165–R190. https://doi. org/10.1152/ajpregu.00163.2017.
- 157. Ibeh, S., Bakkar, N.M.Z., Ahmad, F., Nwaiwu, J., Barsa, C., Mekhjian, S., Reslan, M.A., Eid, A.H., Harati, H., Nabha, S., et al. (2023). High fat diet exacerbates long-term metabolic, neuropathological, and behavioral derangements in an experimental mouse model of traumatic brain injury. Life Sci. *314*, 121316. https://doi.org/10.1016/j.lfs.2022.121316.
- 158. Zhou, M., Du, M., Tang, R., Liu, H.Y., Gao, Z., Wang, Y., You, H.Y., Hao, J.W., Ji, Z.S., Wang, D., and Zhang, Q.H. (2023). Central GLP-1 Resistance Induced by Severe Traumatic Brain Injury Was Associated with Persistent Hyperglycemia in Humans. Neuroendocrinology *113*, 625–640. https://doi.org/10.1159/000529438.
- 159. Li, H., Sun, J., Du, J., Wang, F., Fang, R., Yu, C., Xiong, J., Chen, W., Lu, Z., and Liu, J. (2018). Clostridium butyricum exerts a neuroprotective effect in a mouse model of traumatic brain injury via the gut-brain axis. Neuro Gastroenterol. Motil. *30*, e13260. https://doi.org/10.1111/ nmo.13260.
- 160. Lv, C., Han, S., Sha, Z., Liu, M., Dong, S., Zhang, C., Li, Z., Zhang, K., Lu, S., Xu, Z., et al. (2023). Cerebral glucagon-like peptide-1 receptor activation alleviates traumatic brain injury by glymphatic system regulation in mice. CNS Neurosci. Ther. *29*, 3876–3888. https://doi.org/10.1111/ cns.14308.
- 161. Zhang, J., Yi, T., Cheng, S., and Zhang, S. (2020). Glucagon-like peptide-1 receptor agonist Exendin-4 improves neurological outcomes by attenuating TBI- induced inflammatory responses and MAPK activation in rats. Int. Immunopharm. *86*, 106715. https://doi.org/10.1016/j.intimp.2020.106715.
- 162. Iliff, J.J., Chen, M.J., Plog, B.A., Zeppenfeld, D.M., Soltero, M., Yang, L., Singh, I., Deane, R., and Nedergaard, M. (2014). Impairment of glymphatic pathway function promotes tau pathology after traumatic brain injury. J. Neurosci. 34, 16180-16193. https://doi.org/10.1523/JNEUROS 3020-14.2014.
- 163. Hakon, J., Ruscher, K., Romner, B., and Tomasevic, G. (2015). Preservation of the blood brain barrier and cortical neuronal tissue by liraglutide, a long acting glucagon-like-1 analogue, after experimental traumatic brain injury. PLoS One *10*, e0120074. https://doi.org/10.1371/journal.pone. 0120074.
- 164. Li, Y., Bader, M., Tamargo, I., Rubovitch, V., Tweedie, D., Pick, C.G., and Greig, N.H. (2015). Liraglutide is neurotrophic and neuroprotective in neuronal cultures and mitigates mild traumatic brain injury in mice. J. Neurochem. *135*, 1203–1217. https://doi.org/10.1111/jnc.13169.
- 165. Bader, M., Li, Y., Tweedie, D., Shlobin, N.A., Bernstein, A., Rubovitch, V., Tovar-y-Romo, L.B., DiMarchi, R.D., Hoffer, B.J., Greig, N.H., and Pick, C.G. (2019). Neuroprotective Effects and Treatment Potential of Incretin Mimetics in a Murine Model of Mild Traumatic Brain Injury. Front. Cell Dev. Biol. *7*, 356. https://doi.org/10.3389/fcell.2019.00356.
- 166. Toth, P., Tarantini, S., Csiszar, A., and Ungvari, Z. (2017). Functional vascular contributions to cognitive impairment and dementia: mechanisms and consequences of cerebral autoregulatory dysfunction, endothelial impairment, and neurovascular uncoupling in aging. Am. J. Physiol. Heart Circ. Physiol. *312*, H1–H20. https://doi.org/10.1152/ajpheart. 00581.2016.
- 167. Bouhrara, M., Alisch, J.S.R., Khattar, N., Kim, R.W., Rejimon, A.C., Cortina, L.E., Qian, W., Ferrucci, L., Resnick, S.M., and Spencer, R.G. (2020). Association of cerebral blood flow with myelin content in cognitively unimpaired adults. BMJ Neurol. Open *2*, e000053. https://doi.org/10.1136/ bmjno-2020-000053.
- 168. Gong, Z., Bilgel, M., Kiely, M., Triebswetter, C., Ferrucci, L., Resnick, S.M., Spencer, R.G., and Bouhrara, M. (2023). Lower myelin content is associated with more rapid cognitive decline among cognitively unimpaired individuals. Alzheimers Dement. *19*, 3098–3107. https://doi.org/ 10.1002/alz.12968.
- 169. Bouhrara, M., Khattar, N., Elango, P., Resnick, S.M., Ferrucci, L., and Spencer, R.G. (2021). Evidence of association between obesity and lower cerebral myelin content in cognitively unimpaired adults. Int. J. Obes. *45*, 850–859. https://doi.org/10.1038/s41366-021-00749-x.
- 170. Chen, B., de Launoit, E., Renier, N., and Schneeberger, M. (2024). Central myelin dysfunction bridges obesity and neurological diseases. Trends Endocrinol. Metabol. *35*, 7–10. https://doi.org/10.1016/j.tem.2023. 09.004.

Review

- 171. Langley, M.R., Yoon, H., Kim, H.N., Choi, C.I., Simon, W., Kleppe, L., Lanza, I.R., LeBrasseur, N.K., Matveyenko, A., and Scarisbrick, I.A. (2020). High fat diet consumption results in mitochondrial dysfunction, oxidative stress, and oligodendrocyte loss in the central nervous system. Biochim. Biophys. Acta, Mol. Basis Dis. *1866*, 165630. https://doi.org/10. 1016/j.bbadis.2019.165630.
- 172. Langley, M.R., Choi, C.I., Peclat, T.R., Guo, Y., Simon, W.L., Yoon, H., Kleppe, L., Lucchinetti, C.F., Chini, C.C.S., Chini, E.N., and Scarisbrick, I.A. (2021). Critical Role of Astrocyte NAD⁺ Glycohydrolase in Myelin Injury and Regeneration. J. Neurosci. *41*, 8644–8667. https://doi.org/ 10.1523/JNEUROSCI.2264-20.2021.
- 173. Mojaverrostami, S., Pasbakhsh, P., Madadi, S., Nekoonam, S., Zarini, D., Noori, L., Shiri, E., Salama, M., Zibara, K., and Kashani, I.R. (2020). Calorie restriction promotes remyelination in a Cuprizone-Induced demyelination mouse model of multiple sclerosis. Metab. Brain Dis. *35*, 1211– 1224. https://doi.org/10.1007/s11011-020-00597-0.
- 174. Kaplanis, S.I., Kaffe, D., Ktena, N., Lygeraki, A., Kolliniati, O., Savvaki, M., and Karagogeos, D. (2023). Nicotinamide enhances myelin production after demyelination through reduction of astrogliosis and microgliosis. Front. Cell. Neurosci. *17*, 1201317. https://doi.org/10.3389/fncel.2023. 1201317.
- 175. Cashion, J.M., Young, K.M., and Sutherland, B.A. (2023). How does neurovascular unit dysfunction contribute to multiple sclerosis? Neurobiol. Dis. *178*, 106028. https://doi.org/10.1016/j.nbd.2023.106028.
- 176. Lutfullin, I., Eveslage, M., Bittner, S., Antony, G., Flaskamp, M., Luessi, F., Salmen, A., Gisevius, B., Klotz, L., Korsukewitz, C., et al. (2023). Association of obesity with disease outcome in multiple sclerosis. J. Neurol. Neurosurg. Psychiatry *94*, 57–61. https://doi.org/10.1136/jnnp-2022- 329685.
- 177. Ammar, R.A., Mohamed, A.F., Kamal, M.M., Safar, M.M., and Abdelkader, N.F. (2022). Neuroprotective effect of liraglutide in an experimental mouse model of multiple sclerosis: role of AMPK/SIRT1 signaling and NLRP3 inflammasome. Inflammopharmacology *30*, 919–934. https:// doi.org/10.1007/s10787-022-00956-6.
- 178. Gharagozloo, M., Smith, M.D., Sotirchos, E.S., Jin, J., Meyers, K., Taylor, M., Garton, T., Bannon, R., Lord, H.N., Dawson, T.M., et al. (2021). Therapeutic Potential of a Novel Glucagon-like Peptide-1 Receptor Agonist, NLY01, in Experimental Autoimmune Encephalomyelitis. Neurotherapeutics *18*, 1834–1848. https://doi.org/10.1007/s13311-021-01088-5.
- 179. Lee, C.H., Jeon, S.J., Cho, K.S., Moon, E., Sapkota, A., Jun, H.S., Ryu, J.H., and Choi, J.W. (2018). Activation of Glucagon-Like Peptide-1 Receptor Promotes Neuroprotection in Experimental Autoimmune Encephalomyelitis by Reducing Neuroinflammatory Responses. Mol. Neurobiol. *55*, 3007–3020. https://doi.org/10.1007/s12035-017-0550-2.
- 180. Elbaz, E.M., Senousy, M.A., El-Tanbouly, D.M., and Sayed, R.H. (2018). Neuroprotective effect of linagliptin against cuprizone-induced demyelination and behavioural dysfunction in mice: A pivotal role of AMPK/ SIRT1 and JAK2/STAT3/NF-K B signalling pathway modulation. Toxicol. Appl. Pharmacol. *352*, 153–161. https://doi.org/10.1016/j.taap.2018. 05.035.
- 181. Fultz, N.E., Bonmassar, G., Setsompop, K., Stickgold, R.A., Rosen, B.R., Polimeni, J.R., and Lewis, L.D. (2019). Coupled electrophysiological, hemodynamic, and cerebrospinal fluid oscillations in human sleep. Science *366*, 628–631. https://doi.org/10.1126/science.aax5440.
- 182. Williams, S.D., Setzer, B., Fultz, N.E., Valdiviezo, Z., Tacugue, N., Diamandis, Z., and Lewis, L.D. (2023). Neural activity induced by sensory stimulation can drive large-scale cerebrospinal fluid flow during wakefulness in humans. PLoS Biol. *21*, e3002035. https://doi.org/10.1371/journal.pbio.3002035.
- 183. Holstein-Rønsbo, S., Gan, Y., Giannetto, M.J., Rasmussen, M.K., Si-gurdsson, B., Beinlich, F.R.M., Rose, L., Untiet, V., Hablitz, L.M., Kelley, D.H., and Nedergaard, M. (2023). Glymphatic influx and clearance are accelerated by neurovascular coupling. Nat. Neurosci. *26*, 1042–1053. https://doi.org/10.1038/s41593-023-01327-2.
- 184. Bothwell, S.W., Janigro, D., and Patabendige, A. (2019). Cerebrospinal fluid dynamics and intracranial pressure elevation in neurological diseases. Fluids Barriers CNS *16*, 9. https://doi.org/10.1186/s12987-019- 0129-6.
- 185. Jiang, Q., Zhang, L., Ding, G., Davoodi-Bojd, E., Li, Q., Li, L., Sadry, N., Nedergaard, M., Chopp, M., and Zhang, Z. (2017). Impairment of the glymphatic system after diabetes. J. Cerebr. Blood Flow Metabol. *37*, 1326–1337. https://doi.org/10.1177/0271678X16654702.
- 186. Bao, J., Liang, Z., Gong, X., Yu, J., Xiao, Y., Liu, W., Wang, X., Wang, J.Z., and Shu, X. (2022). High Fat Diet Mediates Amyloid-beta Cleaving Enzyme 1 Phosphorylation and SUMOylation, Enhancing Cognitive Impairment in APP/PS1 Mice. J. Alzheimers Dis. *85*, 863–876. https:// doi.org/10.3233/JAD-215299.
- 187. Eide, P.K., and Hansson, H.A. (2022). A New Perspective on the Pathophysiology of Idiopathic Intracranial Hypertension: Role of the Glia-Neuro-Vascular Interface. Front. Mol. Neurosci. *15*, 900057. https://doi.org/10.3389/fnmol.2022.900057.
- 188. Yiangou, A., Mollan, S.P., and Sinclair, A.J. (2023). Idiopathic intracranial hypertension: a step change in understanding the disease mechanisms. Nat. Rev. Neurol. *19*, 769–785. https://doi.org/10.1038/s41582-023-00893-0.
- 189. Subramaniam, S., and Fletcher, W.A. (2017). Obesity and Weight Loss in Idiopathic Intracranial Hypertension: A Narrative Review. J. Neuro Ophthalmol. *37*, 197–205. https://doi.org/10.1097/WNO.0000000000000448.
- 190. Alperin, N., Ranganathan, S., Bagci, A.M., Adams, D.J., Ertl-Wagner, B., Saraf-Lavi, E., Sklar, E.M., and Lam, B.L. (2013). MRI evidence of impaired CSF homeostasis in obesity-associated idiopathic intracranial hypertension. AJNR. Am. J. Neuroradiol. *34*, 29–34. https://doi.org/10. 3174/ajnr.A3171.
- 191. Alimajstorovic, Z., Pascual-Baixauli, E., Hawkes, C.A., Sharrack, B., Loughlin, A.J., Romero, I.A., and Preston, J.E. (2020). Cerebrospinal fluid dynamics modulation by diet and cytokines in rats. Fluids Barriers CNS *17*, 10. https://doi.org/10.1186/s12987-020-0168-z.
- 192. Westgate, C.S.J., Hagen, S.M., Israelsen, I.M.E., Hamann, S., Jensen, R.H., and Eftekhari, S. (2022). The impact of obesity-related raised intracranial pressure in rodents. Sci. Rep. *12*, 9102. https://doi.org/10.1038/ s41598-022-13181-6.
- 193. Mollan, S.P., Mitchell, J.L., Ottridge, R.S., Aguiar, M., Yiangou, A., Alimajstorovic, Z., Cartwright, D.M., Grech, O., Lavery, G.G., Westgate, C.S.J., et al. (2021). Effectiveness of Bariatric Surgery vs Community Weight Management Intervention for the Treatment of Idiopathic Intracranial Hypertension: A Randomized Clinical Trial. JAMA Neurol. *78*, 678–686. https://doi.org/10.1001/jamaneurol.2021.0659.
- 194. Botfield, H.F., Uldall, M.S., Westgate, C.S.J., Mitchell, J.L., Hagen, S.M., Gonzalez, A.M., Hodson, D.J., Jensen, R.H., and Sinclair, A.J. (2017). A glucagon-like peptide-1 receptor agonist reduces intracranial pressure in a rat model of hydrocephalus. Sci. Transl. Med. *9*, eaan0972. https:// doi.org/10.1126/scitranslmed.aan0972.
- 195. Nelson, R., Silliman, S.L., and Zarroli, K. (2024). Symptomatic idiopathic intracranial hypertension triggered by Ramadan intermittent fasting: a case report. Nutr. Neurosci. *27*, 913–916. https://doi.org/10.1080/ 1028415X.2023.2272090.
- 196. Mitchell, J.L., Lyons, H.S., Walker, J.K., Yiangou, A., Grech, O., Alimaj-storovic, Z., Greig, N.H., Li, Y., Tsermoulas, G., Brock, K., et al. (2023). The effect of GLP-1RA exenatide on idiopathic intracranial hypertension: a randomized clinical trial. Brain *146*, 1821–1830. https://doi.org/10. 1093/brain/awad003.
- 197. Krajnc, N., Itariu, B., Macher, S., Marik, W., Harreiter, J., Michl, M., Novak, K., Wöber, C., Pemp, B., and Bsteh, G. (2023). Treatment with GLP-1 receptor agonists is associated with significant weight loss and favorable headache outcomes in idiopathic intracranial hypertension. J. Headache Pain *24*, 89. https://doi.org/10.1186/s10194-023-01631-z.
- 198. Alimajstorovic, Z., Mitchell, J.L., Yiangou, A., Hancox, T., Southam, A.D., Grech, O., Ottridge, R., Winder, C.L., Tahrani, A.A., Tan, T.M., et al. (2023). Determining the role of novel metabolic pathways in driving intracranial pressure reduction after weight loss. Brain Commun. *5*, fcad272. https://doi.org/10.1093/braincomms/fcad272.
- 199. Grech, O., Mitchell, J.L., Lyons, H.S., Yiangou, A., Thaller, M., Tsermoulas, G., Brock, K., Mollan, S.P., and Sinclair, A.J. (2024). Effect of glucagon like peptide-1 receptor agonist exenatide, used as an intracranial pressure lowering agent, on cognition in Idiopathic Intracranial Hypertension. Eye *38*, 1374–1379. https://doi.org/10.1038/s41433-023- 02908-y.

